MECHANISMS OF VASCULAR DISEASE:

A REFERENCE BOOK FOR VASCULAR SPECIALISTS

Edited by Robert Fitridge and Matthew Thompson Completely Updated Edition 2011

BARR SMITH PRESS

Mechanisms of Vascular Disease

Mechanisms of Vascular Disease: A Reference Book for Vascular Specialists

Robert Fitridge

The University of Adelaide, The Queen Elizabeth Hospital, Woodville, Australia

Matthew Thompson St George's Hospital Medical School, London, UK



BARR SMITH PRESS

An imprint of The University of Adelaide Press

Published in Adelaide by

The University of Adelaide, Barr Smith Press Barr Smith Library The University of Adelaide South Australia 5005 press@adelaide.edu.au www.adelaide.edu.au/press

The University of Adelaide Press publishes peer-reviewed scholarly works by staff via Open Access online editions and print editions.

The Barr Smith Press is an imprint of the University of Adelaide Press, reserved for scholarly works which are not available in Open Access, as well as titles of interest to the University and its associates. The Barr Smith Press logo features a woodcut of the original Barr Smith Library entrance.

© The Contributors 2011

This book is copyright. Apart from any fair dealing for the purposes of private study, research, criticism or review as permitted under the Copyright Act, no part may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without the prior written permission. Address all inquiries to the Director at the above address.

This CIP cataloguing for this work is as follows;

Mechanisms of vascular disease : a reference book for vascular surgeons / Robert Fitridge, Matthew Thompson, [editors].

- 1. Blood vessels, Diseases.
- 2. Blood vessels, Surgery.
- I. Fitridge, Robert
- II. Thompson, M. M.

For the full Cataloguing-in-Publication data please contact National Library of Australia: cip@nla.gov.au

ISBN (paperback) 978-0-9871718-2-5

Book design: Midland Typesetters

Cover design: Emma Spoehr, based on a diagram by Dave Heinrich of the Medical Illustration and Media Unit, Flinders Medical Centre

Paperback edition printed by Griffin Press, South Australia

Table of Contents

Contributors vii Detailed Contents xi

- 1. Endothelium 1 Paul Kerr, Raymond Tam, Frances Plane (Calgary, Canada)
- Vascular smooth muscle structure and function 13 David Wilson (Adelaide, Australia)
- 3. Atherosclerosis 25 Gillian Cockerill, Qingbo Xu (London, UK)
- 4. Mechanisms of plaque rupture 43 Ian Loftus (London, UK)
- Current and emerging therapies in atheroprotection 79 Stephen Nicholls, Rishi Puri (Cleveland, USA)
- Molecular approaches to revascularisation in peripheral vascular disease 103 Greg McMahon, Mark McCarthy (Leicester, UK)
- Biology of restenosis and targets for intervention 115 *Richard Kenagy (Seattle, USA)*
- 8. Vascular arterial haemodynamics 153 Michael Lawrence-Brown, Kurt Liffman, James Semmens, Ilija Sutalo (Melbourne & Perth, Australia)
- 9. Physiological haemostasis 177 Simon McRae (Adelaide, Australia)
- 10. Hypercoagulable states 189 Simon McRae (Adelaide, Australia)
- 11. Platelets in the pathogenesis of vascular disease and their role as a therapeutic

target 201 Sandeep Prabhu, Rahul Sharma, Karlheinz Peter (Melbourne, Australia)

- 12. Pathogenesis of aortic aneurysms 227 Jonathan Golledge, Guo-Ping Shi, Paul Norman (Townsville & Perth, Australia; Boston, USA)
- 13. Pharmacological treatment of aneurysms 247 Matthew Thompson, Janet Powell (London, UK)
- Aortic dissection and connective tissue disorders 255 Mark Hamilton (Adelaide, Australia)
- 15. Biomarkers in vascular disease 277 Ian Nordon, Robert Hinchliffe (London, UK)
- Pathophysiology and principles of management of vasculitis and Raynaud's phenomenon 295 *Martin Veller (Johannesburg, South Africa)*
- 17. SIRS, sepsis and multiorgan failure 315 Vishwanath Biradar, John Moran (Adelaide, Australia)
- Pathophysiology of reperfusion injury 331 Prue Cowled, Robert Fitridge (Adelaide, Australia)
- 19. Compartment syndrome 351 Edward Choke, Robert Sayers, Matthew Bown (Leicester, UK)
- 20. Pathophysiology of pain 375 Stephan Schug, Helen Daly, Kathryn Stannard (Perth, Australia)

- 21. Postamputation pain 389 Stephan Schug, Gail Gillespie (Perth, Australia)
- 22. Treatment of neuropathic pain 401 Stephan Schug, Kathryn Stannard (Perth, Australia)
- 23. Principles of wound healing 423 Gregory Schultz, Gloria Chin, Lyle Moldawer, Robert Diegelmann (Florida, USA)
- 24. Pathophysiology and principles of varicose veins 451 Andrew Bradbury (Birmingham, UK)
- Chronic venous insufficiency and leg ulceration: Principles and vascular biology 459 *Michael Stacey (Perth, Australia)*

- Pathophysiology and principles of management of the diabetic foot 475 David Armstrong, Timothy Fisher, Brian Lepow, Matthew White, Joseph Mills (Tucson, USA)
- Lymphoedema Principles, genetics and pathophysiology 497 *Matt Waltham (London, UK)*
- 28. Graft materials past and future 511 Mital Desai, George Hamilton (London, UK)
- 29. Pathophysiology of vascular graft infections 537 *Mauro Vicaretti (Sydney, Australia)*

Index 549

List of Contributors

David G Armstrong The University of Arizona Southern Arizona Limb Salvage Alliance Tucson, AZ USA

Vishwanath Biradar Intensive Care Unit The Queen Elizabeth Hospital Woodville, SA Australia

Matthew Bown Department of Vascular Surgery University of Leicester Leicester UK

Andrew W Bradbury University Department of Vascular Surgery Birmingham Heartlands Hospital Birmingham UK

Edward Choke Department of Vascular Surgery University of Leicester Leicester UK

Gillian Cockerill Department of Clinical Sciences St George's Hospital Medical School London UK Prue Cowled Department of Surgery University of Adelaide The Queen Elizabeth Hospital Woodville, SA Australia

Helen Daly Royal Perth Hospital Perth, WA Australia

Mital Desai University Department of Vascular Surgery Royal Free Hospital University College London UK

Robert F Diegelmann Department of Biochemistry Medical College of Virginia Richmond, VA USA

Timothy K Fisher Rashid Centre for Diabetes and Research Sheikh Khalifa Hospital Ajmon UAE

Robert A Fitridge Department of Surgery University of Adelaide The Queen Elizabeth Hospital Woodville, SA Australia Gail Gillespie Royal Perth Hospital Perth, WA Australia

Jonathan Golledge Vascular Biology Unit School of Medicine & Dentistry James Cook University Townsville, QLD Australia

George Hamilton University Department of Vascular Surgery Royal Free Hospital University College London UK

Mark Hamilton Department of Surgery University of Adelaide The Queen Elizabeth Hospital Woodville, SA Australia

Robert J Hinchliffe St George's Vascular Institute St George's Hospital London UK

Richard D Kenagy Department of Surgery University of Washington Seattle, WA USA

Paul Kerr Department of Pharmacology University of Alberta Alberta Canada Michael MD Lawrence-Brown Curtin Health Innovation Research Institute Curtin University Perth, WA Australia

Brian Lepow The University of Arizona Department of Surgery Southern Arizona Limb Salvage Alliance Tucson, AZ USA

Kurt Liffman CSIRO Material Science & Engineering and School of Mathematical Sciences Monash University Melbourne, Vic Australia

Ian Loftus Department of Vascular Surgery St George's Hospital London UK

Mark J McCarthy Department of Surgery and Cardiovascular Sciences University of Leicester Leicester UK

Greg S McMahon Department of Surgery and Cardiovascular Sciences University of Leicester Leicester UK

Simon McRae Adult Haemophilia Treatment Centre SA Pathology Adelaide, SA Australia Joseph L Mills The University of Arizona Southern Arizona Limb Salvage Alliance Tucson, AZ USA

Lyle Moldawer Department of Surgery University of Florida Gainesville, FL USA

John L Moran Faculty of Health Sciences University of Adelaide The Queen Elizabeth Hospital Woodville, SA Australia

Stephen Nicholls The Heart and Vascular Institute Cleveland Clinic Cleveland, OH USA

Ian M Nordon St George's Vascular Institute St George's Hospital London UK

Paul E Norman School of Surgery University of WA Fremantle, WA Australia

Karlheinz Peter Baker IDI Heart & Diabetes Institute Melbourne, Vic Australia

Frances Plane Department of Pharmacology University of Alberta Alberta Canada Janet T Powell Imperial College London UK

Sandeep Prabhu Baker IDI Heart & Diabetes Institute Alfred Hospital Melbourne, Vic Australia

Rishi Puri The Heart and Vascular Institute Cleveland Clinic Cleveland, OH USA

Stephan A Schug Royal Perth Hospital Perth, WA Australia

Gregory S Schultz Department of Obstetrics and Gynaecology University of Florida Gainesville, FL USA

Rahul Sharma Baker IDI Heart & Diabetes Institute Alfred Hospital Melbourne, Vic Australia

Guo-Ping Shi Department of Cardiovascular Medicine Brigham & Women's Hospital Harvard Medical School Boston, MA USA

Michael Stacey University Department of Surgery Fremantle Hospital Fremantle, WA Australia Ilija D Sutalo CSIRO Material Science & Engineering and Curtin Health Innovation Research Instutute Curtin University Highett, Vic

Raymond Tam Department of Pharmacology University of Alberta Alberta Canada

Matthew Thompson St Georges Hospital Medical School London UK

Martin Veller Department of Surgery University of Witwatersrand Johannesburg South Africa

Mauro Vicaretti Department of Vascular Surgery Westmead Hospital Westmead, NSW Australia Matt Waltham Academic Department of Surgery St Thomas' Hospital London UK

Matthew L White Vascular and Endovascular Surgery University of Arizona Tucson, AZ USA

David P Wilson School of Medical Sciences Discipline of Physiology University of Adelaide Adelaide SA Australia

Qingbo Xu Department of Cardiology Kings College University of London UK

Detailed Contents

CHAPTER 1 – ENDOTHELIUM

Paul Kerr, Raymond Tam, Frances Plane

Introduction 1 Endothelium-dependent regulation of vascular tone 2 Angiogenesis 7 Haemostasis 8 Inflammation 9 Conclusions 10 References

CHAPTER 2 – VASCULAR SMOOTH MUSCLE STRUCTURE AND FUNCTION

David Wilson

Introduction 13 Smooth muscle (vascular) structure Cytoskeleton 14 Contractile myofilament Functional regulation of vascular smooth muscle: Neuronal, hormonal, receptor mediated 15 Smooth muscle function 17 Myofilament basis of smooth muscle contraction and relaxation Smooth muscle contraction and relaxation 18 Ion channels important in the regulation of smooth muscle function Regulation of cellular Ca²⁺ Sources of cytosolic Ca²⁺ entry 19 Potassium channels Endothelial regulation of smooth muscle vasodilatation 20

Smooth muscle proliferation and vascular remodeling 20 Summary 22 References

CHAPTER 3 – ATHEROSCLEROSIS

Gillian Cockerill, Qingbo Xu

Introduction 25 Atherosclerotic lesions 26 Fatty streaks Plaque or atheroma Hypercholesterolemia and oxidised-LDL 27High-density lipoproteins role in atheroprotection 28 Hypertension and biomechanical stress 29 Biomechanical stress-induced cell death Biomechanical stress and inflammation 31 Biomechanical stress-induced smooth muscle cell proliferation 32 Infections and heat shock proteins Infections Heat shock proteins 33 Infections and HSP expression Infections, sHSP and innate immuntiy 34 Immune responses 36 MHC class II antigens and T cells Oxidised LDL as a candidate antigen HSP60 as a candidate antigen 37 B2-gylcoprotein Ib as a candidate antigen Inflammation

C-reactive protein 38 CD40/CD40L

Summary and perspectives 39 References

CHAPTER 4 – MECHANSIMS OF PLAQUE RUPTURE

Ian Loftus

Introduction 43 Evidence for the 'plaque rupture theory' 44 Coronary circulation Cerebral circulation The role of individual components of the arterial wall The endothelium 45 The lipid core 47 The cap of the plaque 49 Smooth muscle cells and collagen production 50 Macrophages and collagen degradation 51 The vessel lumen 56 The role of angiogenesis in plaque rupture The role of infectious agents in plaque rupture 57 Risk prediction of plaque instability 58 Imaging Blood markers 59 Therapy aimed at plaque stabilisation HMG Co-A reductase inhibitors 60 MMP inhibition Tissue inhibitors of metalloproteinases (TIMPs) 61 Synthetic MMP inhibitors Doxycycline ACE inhibitors Summary 62 References 63

CHAPTER 5 – CURRENT AND EMERGING THERAPIES IN ATHEROPROTECTION

Stephen Nicholls, Rishi Puri

Background 79 Pathology Risk factor modification 80 Statins, LDL lowering and C-reactive protein The complexity of HDL 84 The controversy of trigylcerides 87 Hypertension Risk factor modification in the diabetic patient 89 Glycaemic control Global risk factor reduction in diabetics 91 The metabolic syndrome 92 Future targets 93 Conclusion References 94

CHAPTER 6 – MOLECULAR APPROACHES TO REVASCULARISATION IN PERIPHERAL VASCULAR DISEASE

Greg S McMahon, Mark J McCarthy

Introduction 103 Mechanisms of vascular growth Vasculogenesis Angiogenesis 104 Neovessel maturation 105 Microvascular network maturation 106 Arteriogenesis Therapeutic induction of vascular growth 107 Delivery of molecular activators of vascular growth Angiogenic activators 108 Arteriogenic activators 109 Clinical trials for angiogenic therapy of peripheral vascular disease Conclusions 110 References

CHAPTER 7 – BIOLOGY OF RESTENOSIS AND TARGETS FOR INTERVENTION

Richard Kenagy

Introduction 115 Mechanisms of restenosis Thrombosis 116 Remodelling Intimal hyperplasia 123 Sequence of events after injury Origin of intimal cells 125 Inflammation 126 Role of ECM production 127 The contribution of specific factors to restenosis Growth factors/cytokines Inhibitors 128 Coagulation and fibrinolytic factors 129 Matrix metalloproteinases Extracellular matrix/receptors Targets for intervention 130 Intracellular signalling molecules mTOR and microtubules Transcription factors miRNA 131 Inflammation targets Brachytherapy Extracellular targets and cell-based therapies Angiotensin pathway Cell-based therapies 132 Differential effects on endothelium and SMCs Delivery devices Prevention versus reversal of restenosis Conclusions 133 References 134

CHAPTER 8 – VASCULAR ARTERIAL HAEMODYNAMICS

Michael Lawrence Brown, Kurt Liffman, James Semmens, Ilija Sutalo

Introduction 153

Laplace's law of wall of tension 154 Newtonian fluid 155 Non-Newtonian fluid Poiseuille flow 158 Bernoulli's equation Young's modulus and pulsatile flow 159 Mass conversion 161 Reynold's number Arterial dissection, collateral circulation and competing flows 163 Shear stress and pressure 164 Forces on graft systems 165 Case 1 – The cylindrical graft 168 Case 2 – The windsock graft Case 3 - The curved graft 169 Case 4 – The symmetric bifurcated graft Computational modelling 170 Recent development and future directions 171 Conclusions 172 References 173

CHAPTER 9 – PHYSIOLOGICAL HAEMOSTASIS

Simon McRae

Introduction 177 Primary haemostasis Platelets Platelet adhesion Platelet activation and shape change 179 Platelet aggregation 180 Interactions between primary and secondary haemostasis 181 Secondary haemostasis The coagulation cascade 182 Initiation 183 Amplification Propagation 184 Normal inhibitors of coagulation Fibrinolysis 185 Conclusions 186 References

CHAPTER 10 – HYPERCOAGULABLE STATES

Simon McRae

Introduction 189 Classification of thrombophilia Inherited thrombophilia 190 Type 1 conditions Antithrombin deficiency Protein C and Protein S deficiency Type 2 conditions 191 Factor V Leiden The prothrombin (G20210A) gene mutation FVL/PGM compound heterozygotes Other inherited conditions Acquired thrombophilia 192 Antiphospholipid antibodies Heparin induced thrombocytopenia Myeloproliferative disorders 193 Potential reasons for performing thrombophilia testing Patients with venous thrombosis and their relatives Providing an understanding of the aetiology of a thrombotic event Determining risk of recurrence and therefore optimal duration of anticoagulation 194 Determining the need for primary prophylaxis in asymptomatic family members 195 Making decisions regarding the use of the oral contraceptive pill 196 Determining the need for thromboprophylaxis during pregnancy Patients with arterial thrombosis Potential detrimental effects of thrombophilia testing 197 Conclusion References

CHAPTER 11 – PLATELETS IN THE PATHOGENESIS OF

VASCULAR DISEASE AND THEIR ROLE AS A THERAPEUTIC TARGET

Sandeep Prabhu, Rahul Sharma, Karlheinz Peter

Introduction 201 Platelet function - Adhesion and activation Platelet adhesion 202 Platelet activation 203 Mediators of platelet activation and 'outside in' signalling Thrombin and collagen 204 Adenosine diphosphate (ADP) Thromboxane A2 (TXA2) Adrenaline 206 Second messenger systems 207 Physiological consequences of platelet activation The GP IIb/IIIa receptor and 'insideout' signalling Granule exocytosis 208 Activation-induced conformational change of platelets Platelets and atherosclerosis 209 Role of platelets in the initiation of the atherosclerosis Role of the platelets in the progression of the atherosclerosis Role of platelets in vulnerable plaques and plaque rupture Current and future anti-platelet agents 210Aspirin (salicylic acid) Thienopyridines 211 Clopidogrel Prasugrel 213 Ticlopidine Ticagrelor GPIIb/IIIa Antagonists Other anti-platelet agents and promising new deleopments 214 Platelet function testing 215 Light-transmission aggregometry

Whole blood aggregometry 217 VerifyNow® Assay Flow cytometry 218 **References**

CHAPTER 12 – PATHOGENESIS OF AORTIC ANEURYSMS

Jonathan Golledge, Guo-Ping Shi, Paul E Norman

Introduction 227 Differences between thoracic and abdominal aortic aneurysms 228 Summary of current theories and stages of AAA evolution Atherosclerosis and AAA Immune mechanisms in AAA 229 Extracellular matrix dysfunction 232 Infection 233 **Biomechanical forces** Angiogenesis Intra-luminal thrombus Extracellular matrix proteolysis 234 Genetics 236 AAA rupture 237 Biomechanical factors in aneurysms rupture The role of enzymes in AAA rupture Role of intraluminal thrombus in aneurysm rupture 238 Future research References

CHAPTER 13 – PHARMACOLOGICAL TREATMENT OF ANEURYSMS

Matthew Thompson, Janet T Powell

Background 247 Screening programmes Pathophysiology 248 Therapeutic strategies Beta blockade Modification of the inflammatory response 249 Non-steroidal anti-inflammatories Matrix metalloproteinase (MMP) inhibition Anti-chlamydial therapy 250 Drugs acting on the renin/angiotensin axis HMG Co-A reductase inhibitors 251 The future – Data from recent experimental studies References

CHAPTER 14 – PATHOPHYSIOLOGY OF AORTIC DISSECTION AND CONNECTIVE TISSUE DISORDERS

Mark Hamilton

Introduction 255 Embryology of thoracic aorta and arch vessels Haemodynamics of thoracic compared to abdominal aorta 257 Sizes of normal aorta Classification of aortic syndromes Acute/Chronic DeBakey classification of class 1 dissection – Type 1, 2, and 3 Stanford classification 258 European task force Pathogenesis of thoracic aortic dissection Classical thoracic aortic dissection (class 1 dissection) 260 Intramural haematoma (class 2 aortic dissection) 261 Penetrating aortic ulcer (class 4 aortic dissection) 262 Complications of acute aortic syndromes 263 Visceral ischaemia /malperfusion syndromes Fate of the false lumen Aneurysmal degeneration and rupture 264 Connective tissue disorders and acute

aortic syndromes

xvi

Marfan syndrome Fibrillin and Marfan syndrome 265 The role of transforming growth factor beta in development of the vascular system in health and disease 266 Ehlers-Danlos syndrome 267 Diagnosis of Ehlers-Danlos syndrome 268 Loeys-Deitz syndrome 270 Familial thoracic aortic aneurysm disease 271 Bicuspid aortic valve 273 Turners Syndrome Summary 274 Reference list

CHAPTER 15 – BIOMARKERS IN VASCULAR DISEASE

Ian M Nordon, Robert J Hinchliffe

Introduction 277 What is a biomarker? Types of biomarkers A classical clinical example 278 Potential value of biomarkers in vascular disease 279 Biomarker discovery steps 280 AAA biomarkers Circulating extracellular matrix markers 281 Matrix-degrading enzymes 283 Proteins associated with thrombosis Markers of inflammation 284 Biomarkers of AAA rupture 285 Biomarkers following endovascular repair Inflammation 287 Lipid accumulation Apoptosis Thrombosis Proteolysis 288 Challenges in biomarkers discovery Future work Conclusion 289 References

CHAPTER 16 – PATHOPHYSIOLOGY AND PRINCIPLES OF MANAGEMENT OF VASCULITIS AND RAYNAUD'S PHENOMENON

Martin Veller

Vasculitides 295 Introduction Classification of vasculitides 296 Clinical presentation of vasculitides Investigations of vasculitides Principles of treatment of vasculitides 297 The vasculitides of specific interest to vascular surgeons 298 Giant cell arteritis Takayasu's arteritis 299 Thromboangitis obliterans (Buerger's disease) 300 Behcet's disease 301 Polyarteritis nodosa 302 Vasculitides secondary to connective tissue diseases 303 Systemic lupus erythematosus (SLE) Antiphospholipid antibody syndrome (APS) 304 Rheumatoid arthritis 305 Scleroderma Infective vasculitides 306 Human immunodeficiency virus (HIV) Pathophysiology and principles of Raynaud's phenomenon 307 Prevalence of Raynaud's phenomenon 308 Clinical findings in Raynaud's phenomenon 309 Diagnosis of Raynaud's phenomenon Prognosis 310 Treatment Recommendations 311 References 312

CHAPTER 17 - SIRS, SEPSIS AND

MULTIORGAN FAILURE

Vishwanath Biradar, John Moran

Epidemiology 315 Historical perspectives and definition 316 Risk factors for sepsis 317 Causative agents Pathophysiology of sepsis innate immunity and toll-like receptors (TLRs) 319 Proinflammatory response Coagulation cascade Multiorgan dysfunction syndrome (MODS) 320 Epithelial and endothelial dysfunction Immune suppression and apoptosis Sepsis, circulatory failure and organ dysfunction Management 322 Steroids 323 Recombinant human activated protein C (rhAPC) 324 Glucose control 325 Renal replacement therapy 3-hydroxy-3-methylglutaryl-coenzyme reductase inhibitors (HMG-CoA) 326 Other adjuvant therapies in sepsis Cytokines and anticytokine therapies Pooled immunoglobulin (IVIG) Acute respiratory distress syndrome (ARDS) 327 References

CHAPTER 18 – Pathophysiology of

REPERFUSION INJURY Prue Cowled, Rob Fitridge

Introduction 331 Ischaemia ATP and mitochondrial function Gene expression during ischaemia 332 Reperfusion 333 Reactive oxygen species

Eicosanoids 334 Nitric Oxide 335 Endothelin 336 Cytokines Neutrophil and endothelial interactions 338 Complement activation 340 Tissue destruction 341 Proteases and metalloproteinases Apoptotic cell death during ischaemiareperfusion injury No-reflow phenomenon 342 Therapeutic approaches to IRI Ischaemic preconditioning Ischaemic post-conditioning 343 Conditioning effects of volatile anaesthetics Pharmacological treatments 344 Summary 345 References

CHAPTER 19 – COMPARTMENT SYNDROME

Edward Choke, Robert Sayers, Matthew Bown

Definition 351 Acute limb compartment syndrome Incidence Anatomy/physiology 352 Aetiology/pathophysiology Clinical presentation 354 Investigation 355 Treatment 357 Complication of LCS 359 Outcome 360 Acute abdominal compartment syndrome Incidence 361 Actiology Pathological effects of raised intraabdominal pressure 362 Clinical presentation 363 Investigation Treatment 364 Complications of surgical decompression

xvii

Outcome 367 References 368

CHAPTER 20 – PATHOPHYSIOLOGY OF PAIN

Stephan Schug, Helen Daly, Kathryn Stannard

Introduction 375 Peripheral mechanisms Nociception/transduction Conduction 376 Spinal cord mechanisms Ascending systems 377 Descending control Pain modulation 378 Peripheral sensation Central sensitisation in the dorsal horn Neuropathic pain 379 Mechanisms of neuropathic pain Peripheral mechanisms Spontaneous ectopic discharge Altered gene expression Spared sensory neurons Involvement of the sympathetic nervous system 380 Collateral sprouting Effects of bradykinin Central mechanisms Wind up Central sensitization 381 Central disinhibition Expansion in receptive field size (recuruitment) Immediate early gene expression Anatomical re-organisation of the spinal cord Contribution of glial cells to pain conditions 382 Symptoms of neuropathic pain Stimulus-dependent pain Stimulus-independent pain 383 Sympathetically maintained pain (SMP) Neuropathic pain syndromes

Peripheral neuropathies Central neuropathies 385 References

CHAPTER 21 – POST-AMPUTATION PAIN

Stephan Schug, Gail Gillespie

Introduction 389 Classification and incidence of postamputation pain syndromes Stump pain Phantom sensation 390 Phantom limb pain Pathophysiology of post-amputation pain syndromes Peripheral factors Spinal factors 391 Supraspinal factors Current pathophysiological model of postamputation pain syndromes 392 Prevention of post-amputation pain Perioperative lumbar epidural blockade Peripheral nerve blockade 393 NMDA antagonists Evaluation of the patient with postamputation pain syndromes Examination Therapy of post-amputation pain syndromes 394 Calcitonin Ketamine Analgesic and Co-analgesic compounds Opioids 395 Gabapentin Clonazepam Lidocaine Carbamazepine Tricyclic antidepressants (TCA) Selective serotonin reuptake inhibitors Baclofen Capsaicin Symptomatic treatment of pain components 396 Neuropharmacological therapies

Invasive therapies Electroconvulsive therapy (ECT) Nerve blockade Spinal cord stimulation Implantable intrathecal delivery systems Dorsal root entry zone (DREZ) lesions Psychological therapy 397 Future aims References

CHAPTER 22 – TREATMENT OF NEUROPATHIC PAIN

Stephan Schug, Kathryn Stannard

Introduction 401 Principles of treatment Pharmacological treatment 402 Opioids Recommendations for clinical use of opioids Tramadol Mechanism of action Efficacy 403 Adverse effects Recommendations for clinical use of tramadol in neuropathic pain Antidepressants Tricyclic antidepressants (TCAs) Mechanism of action 404 Adverse effects Selective serotonin re-uptake inhibitors (SSRIs) Serotonin/Noradrenaline reuptake inhibitors (SNRIs) 405 Recommendations for clinical use of antidepressants as analgesics Anticonvulsants Mechanism of action 406 Individual medications Clonazepam Gabapentin Pregabalin 407 Carbamazepine Sodium valproate 408

Phenytoin Lamotrigene Recommendations for clinical use of anticonvulsants as analgesics Local anaesthetics and antiarrhythmics 409 Mechanism of action Lignocaine Mexiletine Recommendations for clinical use of lignocaine and mexiletine in neuropathic pain N-methyl-D-aspartate-receptor antagonists (NMDA) Ketamine 410 Other NMDA antagonists Miscellaneous compounds for systemic use Clonidine Efficacy Baclofen Levodopa 411 Cannabinoids Topical treatments Lignocaine 5% medicated plaster Capsaicin 412 Mechanism of action Efficacy Non-pharmacological therapy Transcutaneous electrical nerve stimulation (TENS) Spinal cord stimulation (SCS) 413 Sympathetic nerve blocks Neurosurgical destructive techniques Cognitive behavious therapy References 414

CHAPTER 23 – PRINCIPLES OF WOUND HEALING

Gregory Schultz, Gloria Chin, Lyle Moldawer, Robert Diegelmann

Introduction 423 Phases of acute wound healing Haemostasis

Inflammation 426 Neutrophils 427 Macrophages 428 Proliferative phase 429 Fibroblast migration 430 Collagen and extracellular matrix production Angiogenesis 431 Granulation 432 Epithelialization Remodelling 433 Summary of acute wound healing 435 Comparison of acute and chronic wounds Normal and pathological responses to injury Biochemical differences in the molecular environments of healing and chronic wounds 436 Biological differences in the response of chronic wound cells to growth factors 439 From bench to bedside Role of endocrine hormones in the regulation of wound healing Molecular basis of chronic non-healing wounds Chronic venous stasis ulcers 441 Pressure ulcers Future concepts for the treatment of chronic wounds 442 Bacterial biofilms in chronic wounds 443 Conclusion 445 References

CHAPTER 24 – PATHOPHYSIOLOGY AND PRINCIPLES OF MANAGEMENT OF VARICOSE VEINS

Andrew Bradbury

Introduction 451 Anatomy Histology 452 Physiology Varicose veins 453 Valvular abnormalities Muscle pump failure 455 Venous recirculation Recurrent varicose veins New varicose veins Persistent varicose veins True recurrent varicose veins 456 Cellular and molecular biology of varicose veins Conclusion 457 References

CHAPTER 25 – CHRONIC VENOUS INSUFFICIENCY AND LEG ULCERATION: PRINCIPLES AND VASCULAR BIOLOGY

Michael Stacey

Definitions 459 Chronic venous insuffiency Leg ulceration Assessment of cause of leg ulceration 460 Epidemiology 461 Pathophysiology Venous abnormality Effect of ambulatory venous hypertension on the tissues in the leg 463 Influence of venous disease on the wound healing process 465 Genetic associations with venous ulceration 466 Assessment of venous function 467 Treatment of venous ulceration Compression therapy Dressings 468 Surgery Prevention of venous ulcer recurrence 470Sclerotherapy and other techniques to obliterate surface and perforating veins Other therapies 471 References

CHAPTER 26 – Pathophysiology and Principles of Management

David Armstrong, Timothy Fisher, Brian Lepow, Matthew White, Joseph Mills

OF THE DIABETIC FOOT

Introduction 475 Pathophysiology of the diabetic foot 476 Neuropathy Structural abnormalities/gait abnormalities Angiopathy 478 Diagnosis History and rapid visual screening Neurological examination 479 Monofilament testing Vibration testing Dermatologic examination 480 Anatomy of occlusive disease - vascular examination Prediction of wound healing: assessment of perfusion 481 Arterial imaging Soft tissue imaging 482 Classification systems 483 Diabetes mellitus foot risk classification University of Texas wound classification system Clinical problems and principles of management 484 Ulceration Epidemiology and risk factors Offloading Non-vascular surgical treatment 485 Class I – Elective 486 Class II - Prophylactic Class III – Curative Class IV – Emergency (urgent) Post-operative management Infections 487 Charcot arthopathy Prevention 490 Conclusion 492 References

CHAPTER 27 – LYMPHOEDEMA – PRINCIPLES, GENETICS AND PATHOPHYSIOLOGY

Matt Waltham

Introduction 497 Classification of lymphoedema Classification of primary lymphoedema 498 The genetics of lymphangiogensis in primary lymphoedema 500 Milroy's disease Lymphoedema – distichiasis syndrome 501 Hypotrichosis – lymphoedema – telangiectasia syndrome 502 Meige disease (primary non-syndromic lymphoedema) Other primary lymphoedema disorders 503 Structure and development of the lymphatic circulation Clinical aspects of lymphoedema 505 Summary References

CHAPTER 28 – GRAFT MATERIALS PAST AND FUTURE

Mital Desai, George Hamilton

The pathophysiology of graft healing 511 The peri-anastomotic area Healing of prosthetic grafts 512 The healing process of the anastomosis Graft porosity and permeability Physical properties of prosthetic materials 514 Tubular compliance Anastomotic compliance mismatch The compliance hypothesis of graft failure Synthetic grafts 515 Newer developments of Dacron grafts Modifications and newer developments of PTFE grafts 517 Polyurethane grafts

Newer developments of polyurethane vascular grafts 518 Biological vascular grafts 519 Newer developments of biological vascular grafts 520 Prosthetic graft modifications Modifications to reduce graft infection Modifications to improve patency 521 Nanocomposite grafts Endothelial cell seeding 522 Single stage seeding Two stage seeding Vascular tissue engineering Non-degradable polymer and cell seeding 523 Bioresorbable and biodegradable polymers Combined bioresorbable and tissue engineered grafts 524 Mechanical conditioning of seeded vascular cells Alternative scaffolds Tissue-engineered grafts 525 Graft materials for aortic endografts 526 The future References 527

CHAPTER 29 – PATHOPHYSIOLOGY OF VASCULAR GRAFT INFECTIONS

Mauro Vicaretti

Introduction 537 Natural history of prosthetic vascular graft infections Mechanism of graft contamination at operation 538 Pathogenesis of graft infections Bacteriology of vascular graft infections Investigations for detection of prosthetic graft infections 539 History and physical examination Laboratory investigations Diagnostic imaging 540 Management of prosthetic graft infections Prevention Reduction of prosthetic vascular graft infection with rifampicin bonded gelatin sealed Dacron 541 Established infection Antibiotic therapy Operative management Conclusion 542 References

Acknowledgements

The Editors gratefully acknowledge the outstanding contributions of each Author involved in this reference book. We would also like to acknowledge the invaluable efforts of Ms Sheona Page who has worked tirelessly on this project. We would also like to thank Prue Cowled PhD and Ms Cayley Wright for their assistance.

Abbreviation List

a1-Pl	a1-protease inhibitor
5-HT	5-Hydroxytryptamine/Serotonin
AAA	Abdominal aortic aneurysm
AAS	Acute aortic syndrome
AAV	Adeno-associated viruses
ACE	Angiotensin converting enzyme
ACS	Acute coronary syndrome
ACS	Abdominal compartment syndrome
ACTH	Adrenocorticotropic hormone
ADAMTS	A disintegrin and metalloproteinase with thrombospondin motifs
ADP	Adenosine diphosphate
AIDS	Acquired immune deficiency syndrome
ALI	Acute lung injury
AMP	Adenosine monophosphate
AMPA	α -amino-3 hydroxy-5-methylisoxazole
ANA	Anti-nuclear antibody
ANCA	Anti-neutrophil cytoplasmic antibody
AOD	Aortic occlusive disease
AP1	Activated protein 1
APC	Activated protein C
APC	Antigen presenting cell
APLAS	Antiphospholipid antibody syndrome
ApoAl	Apolipoprotein Al
ApoE	Apolipoprotein E
APS	Antiphospholipid antibody syndrome
APTT	Activated partial thromboplastin time

ARDS	Acute respiratory distress syndrome
AT	Antithrombin
ATP	Adenosine triphosphate
AVP	Ambulatory venous thrombosis
β2-GPI	β2-glycoprotein Ib
bFGF	Basic fibroblast growth factor
BKCa	Large conductance calcium activated potassium channel
BMPs	Bone morphogenetic proteins
BMS	Bare metal stent
CAD	Coronary artery disease
CaM	Calmodulin
CAM	Cell adhesion molecule
cAMP	Cyclic adenosine monophosphate
ССК	Cholecystokinin
cGMP	Cyclic guanine monophosphate
CD	Cluster of differentiation
CD40L	Cluster of differentiation 40 ligand
CEA	Carotid endarterectomy
CETP	Cholesteryl ester transfer protein
CFD	Computational fluid dynamics
CG	Cationized gelatin
CGRP	Calcitonic gene regulated peptide
CHD	Coronary heart disease
CI	Confidence interval
CIMT	Carotid intimal-media thickness
c-JNK	c-Jun N-terminal kinase
CK-MB	Creatinine kinase (Myocardial specific)
CNCP	Chronic noncancer pain
cNOS	Constitutive nitric oxygen synthase enzyme
COX-1	Cyclooxygenase-1
COX-2	Cyclooxygenase-2
CROW	Charcot restraint orthotic walker
CRRT	Continuous renal replacement therapy

CRPS	Complex regional pain syndromes
	complex regional pair syndromes
СТ	Computational tomography
СТА	Computed tomographic angiography
СТD	Connective tissue disorders
CTGF	Connective tissue growth factor
CYP	Cytochrome P450
CVD	Cardiovascular disease
CVI	Chronic venous insufficiency
DAG	Diacylglycerol
DES	Drug-eluting stent
DRG	Dorsal root ganglion
DNA	Deoxyribonucleic acid
DSA	Digital subtraction arteriography
DTS	Dense tubular system
DVT	Deep vein thrombosis
EC	Endothelial cell
ECM	Extracellular matrix
EDCF	Endothelium-derived contracting factor
EDH	Endothelium-dependent hyperpolarisation
EDS	Ehlers-Danlos syndrome
EET	Epoxyeicosatrienoic acids
ELAM-1	Endothelial-leukocyte adhesion molecule-1
ELG	Endoluminal grafts
ELISA	Enzyme linked immunosorbent assay
Ε _κ	Equilibrium potential
E _M	Membrane potential
eNOS	Endothelial nitric oxide synthase enzyme
EPC	Endothelial progenitor cells
EPCR	Endothelial protein C receptor
ePTFE	Expanded polytetrafluoroethylene
ERK	Extracellular signal-regulated kinase
ESR	Erythrocyte sedimentation rate

ET	Essential thrombocytosis
ET-1	Endothelin 1
EVAR	Endovascular aortic aneurysm repair
EVLA	Endovenous LASER ablation
FDA	Food and drug administration
FDPs	Fibrin degradation products (soluble)
FGF	Fibroblast growth factor
FGF-2	Fibroblast growth factor 2
FMN	Flavin mononucleotide
FVL	Factor V Leiden
GABA	Gamma-aminobutyric acid
GABA B	Gamma-aminobutyric acid subtype B
G-CSF	Granulocyte colony stimulating factor
GMCSF	Granulocyte-macrophage colony stimulating factor
GP	Glycoprotein
GPCR	G-protein coupled receptor
GSV	Great saphenous vein
HDL	High density lipoprotein
HDL-C	High density lipoprotein cholesterol
HIF	Hypoxia inducible factor
HIT	Heparin induced thrombocytopenia
HIV	Human immunodeficiency virus
HLA	Human leukocyte antigen
HMG Co-A	Hydroxymethylglutaryl coenzyme-A
HMW	High molecular weight
HPETE	Hydroperoxyeicosatetraenoic acid
HETE	Hydroxyeicosatetraenoic acids
HR	Hazard ratio
hsCRP	High-sensitive C-reactive protein
HSP	Heat shock protein
HUV	Human umbilical vein
IAH	Intra-abdominal hypertension

xxviii Mechanisms of Vascular Disease

IAP	Intra-abdominal pressure
IAPP	Intra-abdominal perfusion pressure
ICAM-1	Inter-cellular adhesion molecule-1
ICAM-2	Inter-cellular adhesion molecule-2
ICP	Intra-compartmental pressure
ICU	Intensive care unit
IFN	Interferon
IGF-1	Insulin-like growth factor-1
IHD	Ischemic heart disease
IL	Interleukin
IL-1	Interleukin-1
IL-1α	Interleukin-1 alpha
IL1-β	Interleukin-1 beta
IL-6	Interleukin-6
IL-8	Interleukin-8
ILT	Intraluminal thrombus
IKCa	Intermediate conductance calcium-activated potassium channels
IMH	Intramural haematoma
IMP	Inosine monophosphate
iNOS	Inducible nitric oxide synthase enzyme
IP(3)	1,4,5-inositol triphosphate
IRI	Ischemia reperfusion injury
IVIG	Intravenous pooled immunoglobulin
IVUS	Intravascular ultrasound
KGF	Keratinocyte growth factor
KGF-2	Keratinocyte growth factor-2
LAP	Latency associated peptide
LCS	Limb compartment syndrome
LDL	Low density lipoprotein
LDS	Loeys-Dietz syndrome
LLC	Large latent complex
LEC	Lymphatic endothelial cells

LFA-1	Lymphocyte function-associated antigen-1
LO	Lipoxygenase
LOX	Lysyl oxidase
LOPS	Loss of protective sensation
LPA	Lysophosphatidic acid
LPS	Lipopolysaccharide
LTA	Lipoteichoic acid
LTGFBP	Latent TGF binding protein
MAC-1	Macrophage-1 antigen
МАРК	Mitogen activated protein kinase
MCP-1	Monocyte chemoattractant protein-1
M-CSF	Macrophage-colony stimulating factor
MFS	Marfan syndrome
MHC	Major histocompatibility
MI	Myocardial infarction
MIP-1	Macrophage inflammatory protein-1
MLC ₂₀	Myosin light chain ₂₀
MLCK	Myosin light chain kinase
MLCP	Myosin light chain phosphatase
MMP	Matrix metalloproteinase
MODS	Multiple organ dysfunction syndrome
MRA	Magnetic resonance angiography
MRI	Magnetic resonance imaging
mRNA	Messenger RNA
MRSA	Methicillin resistant Staphylococcus aureus
MRSE	Methicillin resistant Staphylococcus epidermidis
MRTA	Magnetic resonance tomographic angiography
MTHFR	Methylenetetrahydrofolate reductase
MT-MMP	Membrane-type MMP
MVPS	Mitral valve prolapse syndrome
NADPH	Nicotinamide adenine dinucleotide phosphate
NGF	Nerve growth factor

Nuclear factor kappa B
Nitinol
Non-junctional perforators
N-methyl-D-aspartate
Number needed to harm
Number needed to treat
Nitric oxide
Nitric oxide synthase enzyme
Non-steroidal anti-inflammatory drug
Neovascularisation
Oestrogen/progesterone contraceptive pill
Osteopontin
Osteoprotegerin
Odds ratio
Oxidised low density lipoprotein
Peripheral arterial disease
Platelet activating factor
Plasminogen activator inhibitor
Plasminogen activator inhibitor-1
Protease activated receptor
Protease activated receptor-1
Protease activated receptor-4
Penetrating aortic ulcer
Protein C
Poly (carbonate-urea) urethane
Percutaneous coronary intervention (angioplasty)
Pulmonary capillary wedge pressure
Platelet-derived growth factor
Platelet-derived growth factor- β
Polydioxanone
Platelet-endothelial cell adhesion molecule-1
Pigment epithelium-derived factor
Paclitaxel-eluting stent

PET	Positron emission tomography
PF4	Platelet factor 4
PGI ₂	Prostacyclin
PGG ₂	Prostaglandin G ₂
PGH ₂	Prostaglandin H ₂
PGEl ₂ /PGl ₂	Prostaglandin I ₂
PGN	Peptidoglycan
PHN	Postherpetic neuropathy
PHZ	Para-anastomotic hyper-compliant zone
РІЗК	Phosphatidylinositol 3-kinase
PIP2	Phosphatidylinositol 4,5-bisphosphate
PLC	Phospholipase C
PLOD	Procollagen lysyl hydroxylase
PMCA	Plasma membrane Ca ²⁺ APTases
PMN	Polymorphonuclear leukocyte
POSS	Polyhedral oligomeric silsesquioxanes
PPAR	Peroxisomal proliferation activating receptor
PPI	Proton pump inhibitor
PRV	Polycythaemia rubra vera
PS	Protein S
PSGL-1	P-selectin glycoprotein ligand-1
PT	Prothombin time
PTCA	Percutaneous coronary angioplasty
PTFE	Polytetrafluoroethylene
PTS	Post-thrombotic syndrome
PUFA	Polyunsaturated fatty acid
PVI	Primary valvular incompetence
rAAA	Ruptured AAA
Rac	Ras activated cell adhesion molecule
RANTES	Regulated upon activation, normal T cell expressed and secreted
RAS	Renin angiotensin system
RCT	Randomised controlled trial

RF	Rheumatoid factor
RFA	Radiofrequency ablation
rhAPC	Recombinant human activated protein C
RNA	Ribonucleic acid
ROS	Reactive oxygen species
RR	Relative risk
RSD	Reflex sympathetic dystrophy
S1P	Sphingosine-1-phosphate
SAPK	Stress-activated protein kinase
SCF	Stem cell factor
SCS	Spinal cord stimulation
ScvO2	Superior vena cava venous oxygen saturation
SDF-1	Stromal-cell-derived factor-1
SERCA	Sarco/endoplasmic reticulum CaATPases
SEP	Serum elastin peptides
SES	Sirolimus-eluting stent
SEPS	Subfascial endoscopic perforator surgery
SFA	Superficial femoral artery
SFJ	Sapheno-femoral junction
SIRS	Systemic inflammatory response syndrome
SKCa	Small conductance calcium-activated potassium channels
SLE	Systemic lupus erythematosus
SMA	Smooth muscle alpha actin
SMC	Smooth muscle cell
SMP	Sympathetically maintained pain
SNARE	Soluble N-ethylmaleimide-sensitive factor activating protein receptors
SNP	Single nucleotide polymorphisms
SNRI	Serotonin/Noradrenaline reuptake inhibitors
SPJ	Sapheno-popliteal junction
SPP	Skin perfusion pressure
SR	Sarcoplasmic reticulum
SSRIs	Selective serotonin re-uptake inhibitors
SSV	Small saphenous vein

SVT	Superficial thrombophlebitis
STIM1	Stromal interacting molecule 1
ΤαCΕ	$TNF\alpha$ converting enzyme
TAAD	Thoracic aortic aneurysm disease
TAD	Thoracic aortic dissection
TAFI	Thrombin-activatable fibrinolysis inhibitor
Tc-99 MDP	Technetium-99 methylene diphosphonate
TCA	Tricyclic antidepressant
ТСС	Total contact cast
TCR	T-cell receptor
TENS	Transcutaneous electrical nerve stimulation
TF	Tissue factor
TFPI	Tissue factor pathway inhibitor
TGF	Transforming growth factor
TGF-α	Transforming growth factor-alpha
TGF-β	Transforming growth factor-beta
TGL	Triglycerides
Th	T helper
TIA	Transient ischemic attack
TIMP	Tissue inhibitors of metalloproteinase
TLR	Toll-like receptors
TNF	Tumour necrosis factor
TNF-α	Tumour necrosis factor-alpha
tPA	Tissue-type plasminogen activator
TRP	Transient receptor potential
TRPC	Transmembrane receptor potential canonical
TRPV1	Transmembrane receptor potential Vanilloid-type
TXA2	Thromboxane A2
uPA	Urokinase
UT	University of Texas
VCAM	Vascular cell adhesion molecule
VCAM-1	Vascular cell adhesion molecule-1
VEGF	Vascular endothelial growth factor

xxxiv Mechanisms of Vascular Disease

VEGF-R	Vascular endothelial growth factor receptor
VIP	Vasoactive intestinal peptide
VLA-1	Very late activating antigen-1
VOCC	Voltage operated calcium channels
VPT	Vibratory perception threshold
VSMC	Vascular smooth muscle cells
VTE	Venous thromboembolism
VV	Varicose veins
vWF	von Willebrand factor
XO	Xanthine oxidase

4 • Mechanisms of Plaque Rupture IAN LOFTUS

St George's Vascular Institute, London, UK.

INTRODUCTION

Atherosclerosis continues to cause considerable morbidity and mortality, particularly in the western world. While risk factors have been clearly identified, their precise roles in early atherogenesis are complex. The early development of the plaque is dependent upon interactions between damaged endothelial cells, vessel wall smooth muscle cells and circulating inflammatory cells mediated by the release of cytokines, growth factors and cell adhesion molecules. Plaque formation may represent a cell-mediated immune phenomenon, with a variety of potential antigenic agents identified. Shear stress and flow considerations also play a part.

Atherosclerosis begins in childhood, but it takes decades for atherosclerosis to evolve into the mature plaques responsible for the onset of ischaemic symptoms. Whilst plaque growth due to smooth muscle cell proliferation, matrix synthesis and lipid accumulation may narrow the arterial lumen and ultimately limit blood flow, uncomplicated atherosclerosis is essentially a benign disease. The final clinical outcome depends on whether a plaque becomes unstable, leading to acute disruption of its surface and exposure of its thrombogenic core to the luminal blood flow. The concept of a 'vulnerable plaque' was initially described in 1990^{1,2} and though this initially gained wide acceptance, many authors now favour the broader concept of a 'vulnerable patient', whereby certain systemic and haematological conditions (e.g. relative hypercaogulability) must also be met before plaque rupture will result in symptomatic thrombosis.³

Mature atherosclerotic plaques are composed of a lipid core that is separated from the vessel lumen by a cap composed of fibrillar collagen. Disruption of this cap exposes the plaque's underlying thrombogenic core to the bloodstream, resulting in thromboembolism. This process of 'plaque rupture' is responsible for the majority of acute coronary syndromes (unstable angina, MI)^{4-6,7} and ischaemic cerebral events (stroke, TIA, amaurosis fugax).⁸⁻¹⁰

Unravelling the complex biochemical and haemodynamic factors leading to plaque rupture is one of the greatest challenges facing contemporary medical research. The vital question in plaque pathogenesis is why, after years of indolent growth, life-threatening disruption and subsequent thrombosis should suddenly occur. Plaque stabilisation may prove to be an important clinical strategy for preventing the development of complications.⁶ Identification of 'vulnerable plaques' (i.e., those most at risk of rupture) and 'vulnerable patients' (i.e. those with predisposition to atherothrombotic occlusion) would allow pharmacotherapy to be targeted more effectively. Furthermore, a greater understanding of the mechanisms involved in plaque rupture will lead to improvements in preventative therapy.

EVIDENCE FOR THE 'PLAQUE RUPTURE' THEORY'

Coronary circulation

Evidence that plaque rupture leads to acute coronary syndromes has been provided from a number of sources. Early pathological studies using post-mortem specimens from fatal cases of acute myocardial infarction have revealed that virtually all cases of coronary thrombosis are related to rupture or fissuring of atheromatous plaques, along with evidence of distal embolisation.^{7,11-13} Angioscopic findings in patients with stable angina have identified smooth atheroma within their coronary arteries, but disrupted irregular atheroma in the arteries of those with unstable angina.^{14,15}

Radiological and histological studies have demonstrated that patients with a plaque morphology consisting of large lipid cores and thin fibrous caps are at increased risk of cardiovascular events.¹⁶⁻¹⁸ In addition, these 'unstable' plaques are not necessarily the ones causing severely stenotic lesions.¹⁹⁻²¹

Cerebral circulation

A similar association between carotid plaque rupture and cerebrovascular events has been shown. In patients undergoing multiple TIAs or stroke progression, microemboli can be detected in the middle cerebral artery by transcranial Doppler.^{10,22} Surface ulceration of carotid plaques seen on ultrasound imaging correlates well with symptoms²³ and echolucent (lipid-rich) plaques are at increased risk of causing future cerebrovascular events.

Early work utilising carotid plaques retrieved at carotid endarterectomy, highlighted the relationship between the presence of thrombus and the clinical status of patients.^{24,25} This supported the theory that ischaemic attacks resulted from embolism rather than reduction in cerebral blood flow, particularly as few strokes occur in watershed areas.²⁶

A number of subsequent studies demonstrated a relationship between the presence of intraplaque haemorrhage and patient symptoms.²⁷ Persson et al found that intraplaque haemorrhage appeared more frequently in symptomatic patients than asymptomatic patients,²⁸ while Lusby suggested a relationship between the onset of neurological symptoms and development of plaque haemorrhage.²⁹ Intraplaque haemorrhage may potentially arise *after* cap rupture, though it now seems most likely that it occurs *prior* to plaque breakdown³⁰ and may play an important role in disruption of the fibrous cap.

The most compelling evidence for an association between carotid plaque rupture and ischaemic cerebral events, is that carotid endarterectomy specimens removed from symptomatic patients are more likely to show histological evidence of rupture, compared to those from asymptomatic patients.^{8,9} Van Damme and colleagues showed that 53% of complicated carotid plaques (intraplaque haemorrhage, haematoma, thrombus or ulceration) were symptomatic with a corresponding neurological deficit, compared to 21% of simple uncomplicated plaques.³¹

THE ROLE OF INDIVIDUAL COMPONENTS OF THE ARTERIAL WALL

A number of intrinsic and extrinsic factors have been identified that determine plaque vulnerability: the size and consistency of the plaque core, the thickness and collagen content of the fibrous cap, and inflammation within the plaque. Further factors such as haemodynamic stress upon the plaque may ultimately contribute to cap disruption.

The evolution of a stable to unstable plaque with cap rupture and thrombosis can be outlined in the following simplistic terms (Figure 4.1): Endothelial damage allows passage of inflammatory cells and LDL into the vessel intima; free radicals are responsible for oxidation of the deposited LDL, and oxidized-LDL promotes cytokine and protease release from macrophages; proteases (in addition to other factors) degrade the fibrous cap causing disruption, allowing exposure of thrombogenic material to the blood; local thrombotic and fibrinolytic activity determine the degree of thrombus progression or dissolution.

Each component contributing to plaque rupture will be discussed in further detail. The relevant processes occur in the endothelium, the lipid core, the fibrous cap and the vessel lumen.

The endothelium

The origin of plaque destabilization can be traced back to endothelial dysfunction, or 'activation'. The endothelium is a single layer of highly specialised cells lining the vessel wall/lumen interface. It plays a vital role in modulating vascular permeability, perfusion, contraction and haemostasis. Leukocytes do not bind to normal endothelium. However, endothelial activation leads to the early surface expression of cell adhesion molecules, including VCAM-1, ICAM-1, E-selectin and P-selectin, which permit leukocyte binding. Many of the known atherosclerosis risk factors (e.g., smoking, hyperlipidaemia, hyperglycaemia, hypertension, hyperhomocysteinaemia) exert their damaging effects by causing endothelial activation. 32-37

Activated endothelial cells express chemoattractant cytokines such as MCP-1, M-CSF, IL-1, IL-6 and TNF- α , as well as cell adhesion molecules. This pro-inflammatory environment, in conjunction with the altered permeability of the dysfunctional endothelium, mediates the migration and entry of leucocytes (mainly monocytes and lymphocytes) into the intima.³⁸⁻⁴⁰

The degree of endothelial dysfunction depends upon the balance between endothelial activation and endothelial 'passivation' (see Figure 4.2). Nitric oxide is the predominant molecule responsible for passivation, and the endothelium acts as an autocrine organ in its production.⁴¹ Nitric oxide is an antioxidant, but has other plaque-stabilizing properties including reducing cell adhesion molecule expression,⁴² platelet aggregation and SMC proliferation. Endothelial nitric oxide synthase, the enzyme responsible for nitric oxide production, is increased in people undergoing regular physical exertion, which may partly explain the benefits of exercise in atherosclerosis prevention.43

Endothelial cells are exposed to 3 different types of mechanical force. Hydrostatic forces (generated by the blood) and circumferential stress (generated by the vessel wall) are responsible for endothelial injury and activation. The third force is haemodynamic shear stress (generated by the flow of blood), which is inversely related to atherosclerosis formation - areas of high shear stress being relatively protected.⁴⁴ Despite the systemic nature of atherosclerosis, it is an anatomically focal disease with certain sites having a propensity for plaque formation. Arterial bifurcations exhibit slow blood flow, sometimes even bi-directional flow, resulting in decreased shear stress. The activity of endothelial nitric oxide synthase is decreased in these areas of non-laminar blood flow.45,46 In addition, there is increased oscillatory and turbulent shear stress at bifurcations,

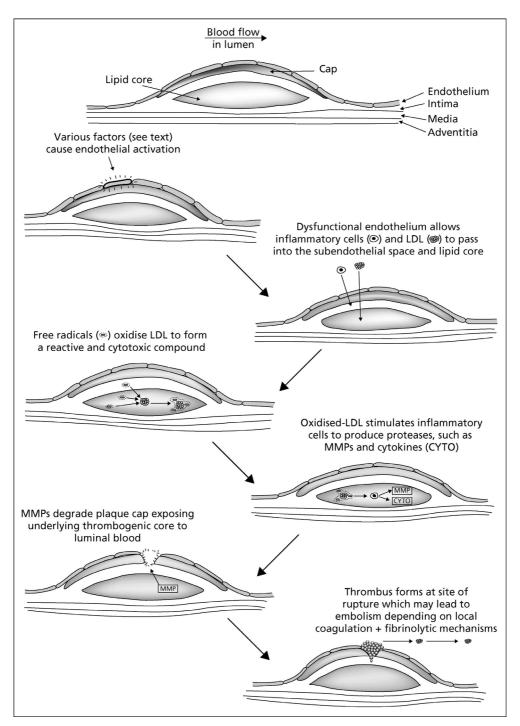


FIGURE 4.1: The stages of plaque rupture.

Endothelial activation	vs Endothelial passivation
 LDL-cholesterol Smoking Homocysteine Glucose Mechanical forces Oxygen free radicals Angiotensin II 	 Nitric oxide Endothelial nitric oxide synthase HMG Co-A reductase inhibitors ACE inhibitors Angiotensin II receptor blockers Polyunsaturated fatty acids

Factors affecting endothelial activity

FIGURE 4.2: Factors affecting endothelial activity.

associated with an increase in oxygen free radical production⁴⁷ and monocyte adhesion.⁴⁸

According to Laplace's law, the higher the blood pressure and the larger the luminal diameter, the more circumferential tension develops in the wall.⁴⁹ This phenomenon combined with a radial compression of the vessel wall may lead to excessive stress in vulnerable regions of the plaque, particularly the cap and shoulder.⁵⁰ For fibrous caps of the same tensile strength, those caps covering moderately stenotic plaques are probably more prone to rupture than those covering severely stenotic plaques, because the former have to bear a greater circumferential tension.⁵¹

The propagating pulse wave causes cyclic changes in lumen size and shape with deformation and bending of plaques, particularly those with a large soft plaque core. Eccentric plaques typically bend at the junction between the relatively stiff plaque and the compliant vessel wall.⁵² The force applied to this region is accentuated by changes in vascular tone.

High blood velocity within stenotic lesions may shear the endothelium away, but whether high wall stress alone may disrupt a stenotic plaque is questionable.⁴ The absolute stresses induced by wall shear are usually much smaller than the mechanical stresses imposed by blood and pulse pressure.⁵³

It is clear that the endothelium is much more than an inert arterial wall lining. It is, in fact, a dynamic autocrine and paracrine organ responsible for the functional regulation of local haemodynamics. Factors that disturb this delicate balance are responsible for the initiation of a cascade of events eventually leading to plaque rupture.

The lipid core

The size and consistency of the atheromatous core is variable and critical to the stability of individual lesions, with a large volume lipid core being one of the constituents of the vulnerable plaque (Figure 4.3). It appears that the accumulation of lipids in the intima renders the plaque inherently unstable.

Although extremely variable, the 'average' coronary plaque is predominantly sclerotic with the atheromatous core making up <30% of the plaque volume.54 The variability in plaque composition is poorly understood, with no relationship to any of the identified risk factors for atherosclerosis. Gertz and Roberts examined the histological composition of post-mortem plaques from coronary arteries.55 17 infarct-related



FIGURE 4.3: Longitudinal section of carotid plaque demonstrating a large volume lipid core

They found much larger proportions of the disrupted plaques to be occupied by atheromatous gruel in comparison to the intact plaques. Davies found a similar relationship in aortic lesions, with 91% of thrombosing plaques versus 11% of intact plaques exhibiting a lipid core that occupied >40% of the total plaque volume.⁵⁶

Histological data regarding the necrotic core of carotid plaques is limited. There is, however, considerable evidence to link ultrasound-detected echolucent plaques (deemed to contain more soft or amorphous tissue) with symptomatology.^{57,58} Feeley and colleagues demonstrated that symptomatic carotid plaques contained a significantly higher proportion of amorphous material than asymptomatic plaques,⁵⁹ with the lipid-rich core constituting 40% of overall plaque volume.⁶⁰

LDL plays a more complex role in plaque instability than can be explained simply by the 'space-occupying' effect of accumulated lipid. A large core may produce a greater luminal narrowing, but plaque rupture sites are often characterized by 'outward remodelling' whereas those stenoses causing stable angina are more likely to be associated with 'inward remodelling'.⁶¹ Indeed, it has been shown that in patients suffering acute coronary syndromes who had undergone angiography in the preceding months, the responsible lesion was recorded as causing a <70% stenosis in the majority of cases.^{19,20,61} This is perhaps not surprising since, as mentioned earlier, a larger lumen places increased circumferential stress on the plaque, predisposing it to rupture.

inflammatory cells As cross the dysfunctional endothelium, cholesterol also enters in the form of LDL, and becomes trapped in the subendothelial space. This LDL is oxidized by free radicals creating a pro-inflammatory compound.62 Oxidized-LDL is taken up by intimal macrophages - the process being mediated via receptors expressed on the macrophage surface,63 although endocytosis of native LDL has also been demonstrated.⁶⁴ This process initially protects the surrounding smooth muscle and endothelial cells from the direct cytotoxic effects of oxidised-LDL, but leads to the formation of 'foam cells' (lipid-laden macrophages). Uptake of oxidized-LDL stimulates the expression of cytokines and proteolytic enzymes, propagating the cycle of inflammation.

The formation of a lipid core is a balance between LDL deposition of cholesterol in the damaged intima and removal by HDL (Figure 4.4). HDL and its carrier, apolipoprotein A-I, are responsible for so-called 'reverse cholesterol transport' moving cholesterol from cells into the blood (from where it can be transferred to the liver for excretion in the bile).65 However, it may also be capable of effecting lipid removal directly from the plaque, one of the possible explanations for plaque regression seen with increased HDL levels.⁶⁶ HDL may have other beneficial effects also, such as improving endothelial function,⁶⁷ decreasing cell adhesion molecule expression,68 and inhibiting oxidation of LDL.69

In addition to the potential proinflammatory role of oxidised LDL, it has recently been proposed that cholesterol accumulation may lead to plaque rupture via a more direct physical pathway. Changes

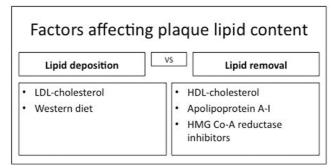


FIGURE 4.4: Factors affecting plaque lipid content

in local biological milieu such as decreased temperature or increased pH may cause in vitro precipitation of cholesterol into solid crystals. This alteration in state not only leads to a significant volume increase of up to 45%, but also leads to formation of sharptipped crystals that might be capable of damaging surrounding tissues and initiating plaque rupture. Using electron microscopy, Abela et al demonstrated that such crystal could be seen perforating the luminal surface of ruptured plaques from human coronary arteries.^{70,71}

The cap of the plaque

The cap of the atherosclerotic plaque plays a vital role in isolating the plaque's thrombogenic core from the bloodstream. Since the thickness and collagen content of this cap are important determinants of overall plaque stability,⁵¹ many authors now use the term 'thin-cap fibroatheroma' (TCFA) to identify those plaques most at risk of rupture. The accepted definition of TCFA is any plaque with a cap thickness of less than 65µm. Though the exact mechanisms that underlie progression from stable plaque to TCFA remain somewhat uncertain, it has been suggested that endothelial shear stress may play an important role, since TCFAs most often arise at sites of low endothelial shear stress (such as bifurcations and the concave side of arterial bends).⁷²

Whatever the thickness of the fibrous cap, it is composed largely of fibrillar collagens (type I and type III⁶⁸), though the relative proportion of collagen decreases as the cap thins. The fibrillar collagens have a lower thrombogenicity than the underlying core, but their exposure can be responsible for thrombus formation following erosion of the overlying endothelium.^{73,74} This phenomenon accounts for one-third of acute coronary syndromes,⁷⁵ and the subsequent healing process of erosions can account for rapid and step-wise progression in plaque growth, leading to sudden increases in stenosis or occlusion.⁷⁶

The most vulnerable area of the plaque is the shoulder region, where the cap is often at its thinnest.7 Studies have shown a reduction in the collagen content of the cap around areas of plaque disruption, as well as steep transverse gradients of connective tissue constituents across ulcerated plaques.77 This may result from a reduction in matrix production by smooth muscle cells, which exhibit diminished numbers in areas of plaque disruption,⁵⁶ or from increased degradation of matrix by proteolytic enzymes. It is most likely, of course, that a combination of excessive matrix degradation and reduced matrix production are responsible for cap thinning (Figure 4.5). A reduction in SMCs within the fibrous cap would certainly undermine its strength.⁷⁸ Recently there has been interest in the role of smooth muscle cell

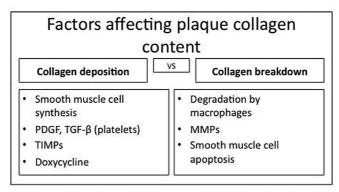


FIGURE 4.5: Factors affecting plaque collagen content

apoptosis in plaque cap weakening, caused by a combination of intrinsic and extrinsic factors, particularly macrophage and lipid derived products.^{79,80}

More recently it has been suggested that plaque integrity may also be influenced by the development of minute, spherical microcalcifications within the fibrous cap. These microcalcifications are thought to represent accumulations of calcified macrophages or even post-apoptotic smooth muscle cells and result in highly focal increases in physical stress. The increased stress leads to areas of facial debonding, weakening the infrastructure of the cap and contributing to subsequent plaque disruption.⁸¹

Smooth muscle cells and collagen production

The SMC has a paradoxical role in plaque instability. On the one hand, SMCs are responsible for plaque matrix production and adverse arterial remodelling, while on the other, they produce collagens that give the plaque intrinsic strength. SMC inhibition therefore has potentially detrimental and beneficial effects.

In the normal arterial wall, SMCs are present in the media and express a differentiated phenotype. They are contractile and do not divide or migrate.⁸² In atherosclerosis, when stimulated by the milieu of growth factors and cyokines, they 'dedifferentiate' and express a synthetic phenotype.⁸³ In the media, SMCs are surrounded by a basal lamina consisting of type IV collagen. Proteolytic enzymes secreted by macrophages are responsible for digestion of this supporting framework. The released SMCs are then able to migrate to the intima, where they secrete new extracellular matrix.84 SMCs play a crucial role in stabilising atherosclerotic plaques, as they are responsible for the production of the cap fibrillar collagens.⁸² In this respect, SMCs are important not only in initial formation of the fibrotic cap, but also in repair of subclinical plaque rupture. SMCs accumulate at the rupture site and secrete fibrous proteins. This restores plaque integrity, but may also lead to rapid growth of the plaque causing vessel stenosis. Certain platelet factors, including PDGF and TGF- β , are felt to be particularly important in stimulating collagen synthesis by SMCs, whereas γ -interferon (from activated T-cells) has the opposite effect.⁸⁵

Since SMCs are the only cells producing fibrous tissue for inclusion in the atherosclerotic plaque, the balance between recruitment and degradation of these cells is clearly of great significance in plaque stability. It had previously been accepted that all SMCs involved in atherosclerosis were derived from the local vessel media or intima. However, many groups are now examining the possibility that they may also be recruited from a circulating pool of SMC progenitor cells.⁸⁶ The prospect of manipulating the activity of these progenitor cells to increase plaque stability is an attractive therapeutic target, though further work is still required in this area.

Whatever the true origin of plaque SMCs, they play a vital role in maintaining the structure of the plaque and SMC apoptosis leads to decreased collagen production, thinning of the fibrous cap and increased volume of the necrotic core.87-89 A recent, though small, study demonstrated that the proportion of SMCs undergoing apoptosis and the frequency of cytoplasmic remnants of apoptotic cells were significantly increased in unstable versus stable angina atherectomy specimens.⁹⁰ Apoptosis of SMCs and macrophages has been identified within plaques, but only in advanced disease with dense macrophage infiltration. Apoptotic cells are deemed to have become susceptible to a form of cell death which is distinct from necrosis and is characterised by a series of morphological changes, starting with shrinkage of the cell membrane and leading on to condensation of nuclear chromatin, cellular fragmentation and eventually engulfment of apoptotic bodies by surrounding cells.⁷⁹

Pro-apoptotic proteins are present in advanced plaques, and it has been observed that cells derived from the plaque, but not the adjacent media, die when brought into culture.^{80,91} Intimal cell apoptosis may account for the low density of smooth muscle cells in unstable plaques, and may contribute to the events leading up to plaque disruption. Though further study is still required, prevention of smooth muscle cell apoptosis may prove to be an important therapeutic target in the treatment of atherosclerotic disease.

Macrophages and collagen degradation

It is now known that inflammation plays a major role in plaque progression and especially in the period just prior to its rupture.⁹² Macrophages control many of the inflammatory processes within the plaque,⁹³ and are responsible for the production of proteolytic enzymes capable of degrading the extracellular matrix.^{94,95} The predominant proteolytic enzymes involved in plaque disruption are the matrix metalloproteinases or MMPs.⁹⁶

The MMPs are a family of proteolytic enzymes characterised by the presence of zinc ions at their active sites. All degrade components of the extracellular matrix, and are divided into 4 main classes on the basis of their substrate specificity (Table 4.1).

MMPs are essential in normal healthy individuals, playing a key role in processes such as wound healing.97,98 However there is growing interest in their role in disease states where ECM breakdown plays a predominant role.99 Early interest focused on a pathological role for MMPs in the resorption of periodontal structures in periodontal disease,100 the destruction of joints in rheumatoid arthritis,¹⁰¹ and the local invasive behaviour of malignancies.¹⁰² In vascular disease, they have been implicated in many of the stages of atherosclerosis but most particularly in acute plaque disruption.¹⁰³ The site of rupture is characterised by an intense inflammatory infiltrate consisting predominantly of macrophages,94 that undergoes activation resulting in increased MMP expression. This shifts the delicate equilibrium towards proteolysis and away from matrix accumulation, making plaque disruption more likely (Figure 4.5).

MMP activity is tightly controlled at several levels and expression of MMPs is determined at the transcriptional level by various cytokines and growth factors.¹⁰⁴

MMP	Alternative names	Principal substrates
Collagenases MMP-1 MMP-8 MMP-13 MMP-18	Collagenase-1, Interstitial collagenase Collagenase-2, Neutrophil collagenase Collagenase-3 Collagenase-4, Xenopus collagenase	Collagens I,II,III, gelatin, MMP-2 & 9 Collagens I,II,III, gelatin Collagens I,II,III, gelatin, PAI-2 Collagen I
Gelatinases MMP-2 MMP-9	Gelatinase-A, 72 kDa gelatinase Gelatinase-B, 92 kDa gelatinase	Gelatin, collagens IV,V,VII,X,XI,XIV, elastin, fibronectin, aggrecan Gelatin, collagen types IV,V,VII,X, elastin
Stromelysins MMP-3 MMP-10 MMP-11	Stromelysin-1 Stromelysin-2 Stromelysin-3	Collagens III,IV,IX,X, gelatin, aggrecan, MMP-1,7,8,9 & 13 Collagens III,IV,V, gelatin, MMP-1 & 8
Matrilysins MMP-7 MMP-26	Matrilysin-1, Pump-1 Matrilysin-2, Endometase	
Membrane types MMP-14 MMP-15 MMP-16 MMP-17 MMP-24 MMP-25	MT1-MMP MT2-MMP MT3-MMP MT4-MMP MT5-MMP MT6-MMP	Collagens I,II,III, gelatin, MMP-2 & 13 MMP-2, gelatin MMP-2
Others MMP-12 MMP-19 MMP-20 MMP-21 MMP-23 MMP-27 MMP-28	Macrophage elastase No trivial name Enamelysin XMMP (Xenopus) Epilysin	

TABLE 4.1: THE MATRIX METALLOPROTEINASE FAMILY

In a variety of tissue types, IL-1, PDGF and TNF- α stimulate expression,^{105,106} while heparin, TGF- β and corticosteroids inhibit expression.^{107,108} In recent years, there has also been considerable interest in the regulatory role of extracellular matrix metalloproteinase inducer (EMMPRIN). EMMPRIN was initially identified as a tumour-derived protein that facilitated cancer cell invasion by stimulating MMP production in epithelial cells and fibroblasts.¹⁰⁹ However, subsequent studies have demonstrated that EMMPRIN also stimulates production of MMPs by smooth muscle cells and monocytes, making it highly relevant in atherosclerosis and plaque instability. In addition, EMMPRIN may also lead to increased production of inflammatory cytokines which further augment MMP activity as described above.¹¹⁰

MMPs are initially secreted as latent inactive proenzymes and converted to the active state by cleavage of a propeptide domain.¹¹¹ The major physiological activator is plasmin, which in turn is regulated by PAI.¹¹² Thrombin has been shown to activate MMP-2 in vitro¹¹³ and could provide a mechanism for MMP activation at sites of vascular injury. Reactive oxygen species also modulate enzyme activation.^{114,115}

Metalloproteinase activity is further governed by naturally occurring MMP inhibitors. These 'tissue inhibitors of metalloproteinases' (TIMPs) provides a further level of control and overall proteolytic activity depends on the ratio of activated MMPs to TIMPs.¹¹⁶

Early studies showed that MMPs were present at increased levels in atherosclerotic arteries. Raised levels of gelatinase activity were demonstrated in the aortas of patients with occlusive disease compared to healthy controls, and zymography revealed that this was predominantly MMP-9.117 Subsequently, quantitative studies using ELISA revealed a six-fold increase in MMP-9 levels in atherosclerotic aortas.118 The level and expression of MMP-2 is also increased in atherosclerotic aortic tissue compared with normal aorta.¹¹⁹ While expression of MMP-2 has been detected in normal arteries, it appears that most MMPs are expressed only in atherosclerotic tissue.¹²⁰ The colocalisation of MMP-1, -2, -3 and -9 to the vulnerable shoulder of the plaque provided further evidence of their potential role in acute disruption.120

More recent studies have demonstrated an association between MMP levels and markers of plaque instability. Increased immunostaining for MMP-9 was seen in 12 atherectomy specimens retrieved from patients with unstable angina compared to the stable form.¹²¹ A larger study, involving 75 carotid endarterectomy specimens, demonstrated a close association between raised plaque levels of MMP-9 and a number of indicators of plaque instability, including symptomatology, cerebral embolisation and histological features of plaque rupture.⁹

Convincing evidence therefore exists of increased levels of MMP-2 and -9 in unstable plaques. However, intact type I and type III collagen molecules, which account for the load-bearing strength of the plaque cap, are not substrates for MMP-2 and -9. While it has been reported that high concentrations of MMP-2 can degrade type I collagen in an in vitro environment devoid of TIMPs,¹²² it is likely that in vivo only the collagenases, MMP-1, -8 and -13, are capable of degrading fibrillar collagens.

MMP-1 and -13 levels are higher in 'atheromatous' compared to 'fibrous' plaques,¹²³ and MMP-8 has been demonstrated in atheroma but not normal arteries.¹²⁴ The expression of MMP-1 is increased in areas of high circumferential stress.¹²⁵ It is likely that both mechanics and proteolysis play a role in the degradation and weakening of the collagen-rich extracellular matrix, and understanding their interaction may be crucial.¹²⁶

Evidence from our laboratories suggests that active MMP-8 is significantly raised in unstable plaques retrieved at carotid endarterectomy (Figure 4.6). The ratio of active MMP-8 to TIMP-1 and -2 (its naturally occurring inhibitors) were also significantly higher in the more unstable plaques of the 159 specimens collected in this study. This implies net proteolysis of the types of collagen found in the cap of the plaque by MMP-8. Immunohistochemistry confirmed the presence of MMP-8 protein within the plaque, which colocalised with macrophages (Figure 4.7). Genetic variation in the genes controlling MMPs could theoretically be responsible for the susceptibility of some individuals to atherosclerotic plaque rupture. Early work has identified a number of polymorphisms that may be influential in this regard. Price et al have identified a novel genetic variation in the MMP-2 gene.¹²⁷ Ye and colleagues detected a polymorphism in the promoter region of the MMP-3 gene that may lead to increased systemic levels.¹²⁸ This polymorphism was subsequently found to be more common in patients suffering MI, compared to a control group.¹²⁹ A single nucleotide polymorphism

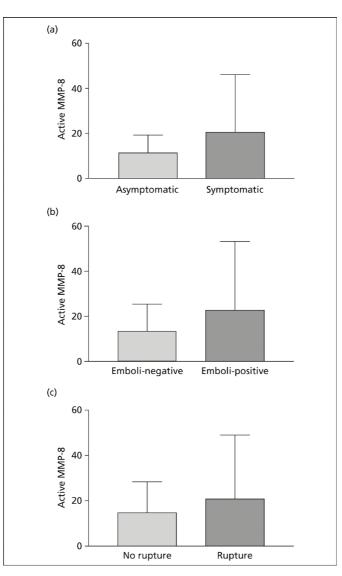
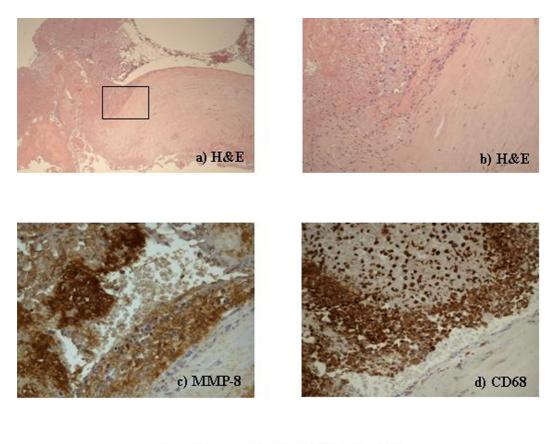


FIGURE 4.6: Plaque concentrations of active MMP-8 are significantly higher in symptomatic compared to asymptomatic carotid plaques: **(a)** from patients suffering carotid territory symptoms in the 6 months prior to surgery (p-value 0.0002), **(b)** from patients with pre-operative cerebral embolisation detected by transcranial Doppler (p-value 0.003) and **(c)** showing histological evidence of plaque rupture. Median values and interquartile ranges shown (p-value 0.003).



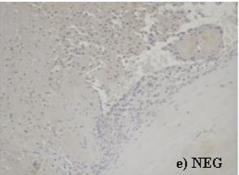


FIGURE 4.7: Histological sections taken from the shoulder region of a symptomatic carotid plaque. Some sections show disruption of the friable plaque.

- (a) Low power H&E section with boxed area delineating high power view shown in (b-e).
- (b) High power H&E section demonstrating a cellular infiltrate.
- (c) Strong reactivity for MMP-8 in cells.
- (d) Positive staining for CD68 (macrophages).
- (e) Negative immunohistochemistry control.

(C to T transition at position –1562) has been shown to influence MMP-9 transcription.¹³⁰ In this study by Zhang and co-workers, triple-vessel coronary artery disease was detected by angiography in 26% of patients with this polymorphism compared to 15% of those without.¹³⁰ Presenting a coherent picture of the interactions between various polymorphisms and the corresponding gene expression is difficult, and further complicated by environmental effects. However, it is clear that the potential exists to identify 'at risk' individuals in such a manner.

The vessel lumen

Disruption alone would not precipitate ischaemic syndromes without thrombus formation on the plaque surface, so plaque instability and thrombogenicity in tandem predispose to acute clinical events. Platelet adherence to the sub-endothelium after surface disruption leads to activation, with ADP and serotonin release stimulating further platelet recruitment and activation.

Once formed, thrombus can behave in three ways, dependent on the physical nature of the rupture and the balance between local fibrinolytic and coagulation processes. Firstly, the initial thrombus may progress to cause occlusion of the vessel. Secondly, the thrombus may disintegrate resulting in distal embolisation. Thirdly, the clot can undergo rapid dissolution, with the healed rupture resulting in a variable decrease in vessel lumen diameter.⁷⁶

Tissue factor is a major regulator of haemostasis.¹³¹ It is the most thrombogenic component of atherosclerotic plaques¹³² and is expressed by numerous cell types, including endothelial cells. The level of tissue factor in coronary plaques from patients with unstable angina is more than twice the value observed in those plaques from stable angina patients.¹³³ Positive immunostaining for

tissue factor correlates with areas of intense macrophageinfiltration and SMCs, suggesting a cell-mediated increased thrombogenicity in unstable plaques. The increase in tissue factor levels seems to be linked to expression of the CD-40 receptor on the macrophage cell surface. The CD-40 ligand is expressed on activated T-lymphocytes, and other atheroma-associated cells,134 which can therefore induce tissue factor production by macrophages via this signalling system. Expression is also regulated by cytokines and oxidised LDL.135,136 It has been reported that a blood-borne pool of tissue factor exists,¹³⁷ though in the context of plaque disruption, macrophage production of tissue factor is predominantly responsible for plaque thrombogenicity.^{133,138,139} It is interesting to note that many of the recognised cardiovascular risk factors increase the expression of tissue factor.140,141

THE ROLE OF ANGIOGENESIS IN PLAQUE RUPTURE

Angiogenesis is essential for normal growth and development. Neovascularisation has been observed in plaques¹⁴² and it is postulated that it may play a role in atherosclerosis by providing growth factors and cytokines to regions of plaque development.

A study of coronary atherectomy specimens revealed the presence of neovascularisation in 50% of specimens from patients with unstable angina compared to 10% of specimens from patients with stable angina,²⁹ suggesting a possible role in plaque instability. Angiogenesis may contribute to plaque instability by causing intraplaque haemorrhage or extravasation of erythrocytes and inflammatory mediators into the centre of the plaque. Once red blood cells have leaked into the plaque, cholesterol from the cell membrane may become incorporated into the lipid core increasing its volume.¹⁴³ This is supported by the finding that lipidrich plaques have a significantly higher microvessel density than fibrous plaques.¹⁴⁴ The associated delivery of inflammatory cells may also lead to plaque degradation by stimulating MMP activity as described earlier in this chapter.

Perhaps more importantly, most neovascularisation occurs at the vulnerable shoulder area of the plaque. Immunostaining for inflammatory cells showed a close association between angiogenesis and inflammatory infiltration. In addition, a parallel increase in the expression of leukocyte adhesion molecules in the same vulnerable areas was demonstrated.¹⁴⁴

Angiogenesis involves interactions between endothelial cells and components of the basement membrane matrix. MMP activity is required for such interactions, especially MMP-2 and MT1-MMP.¹⁴⁵ TIMPs have been shown to reduce angiogenesis, while up-regulation of MMP activity stimulates its increase.¹⁴⁶However, whilst neovascularisation may promote and sustain inflammatory infiltration, the converse may also be true, whereby changes in the plaque associated with inflammation may themselves promote angiogenesis. Further work in this area is required.

THE ROLE OF INFECTIOUS AGENTS IN PLAQUE RUPTURE

The role of infectious agents in atherosclerosis and plaque rupture is controversial. Definitive proof of a causal relationship is lacking, although studies have reported associations between plaque development and *Chlamydia pneumoniae*,¹⁴⁷⁻¹⁴⁹ *Helicobacter pylori*,¹⁵⁰ cytomegalovirus,^{151,152} Herpes simplex virus types 1 and 2,¹⁵³ and hepatitis A virus.¹⁵⁴

Certain infectious agents can evoke cellular and molecular changes supportive of a role in atherogenesis.¹⁵⁵ Work has shown

that Chlamydial interaction with monocytes results in upregulation of TNF- α and IL-1 β ,^{156,157} both of which are associated with plaque development. Chlamydial production of the HSP-60 antigen activates human vascular endothelium, and increases TNF- α and MMP expression in macrophages.^{158,159} Once again, these are factors that influence plaque stability.

It has also been proposed that infective pathogens may exert their effects via direct infection of cells in the vessel wall. This establishes localised inflammation, leading to increased smooth muscle cell migration and greater uptake of oxidised low-density lipoprotein.¹⁶⁰

There is some doubt about the methods employed for Chlamydia detection,¹⁶¹ and also the role of potential confounding factors in epidemiological studies.¹⁶² A large-scale prospective study of 15,000 healthy men in the United States which was controlled for age, smoking, socio-economic status and other cardiovascular risk factors, failed to show any association between Chlamydia seropositivity and the risk of MI.¹⁶³

More recently, the STAMINA trial¹⁶⁴ demonstrated that eradication therapy (amoxicillin/ azithromycin, metronidazole and omeprazole) administered for 1-week after an acute coronary syndrome, significantly reduced cardiac death and acute coronary syndrome readmission rates over the following 12 months. These effects were unrelated to *Chlamydia pneumoniae* or *Helicobacter pylori* seropositivity, however, suggesting that the trial therapy prevented lesion progression by a mechanism unrelated to its antibiotic action.

Though the role of infection in atherosclerosis is still unclear, it seems that any causal relationship is likely to be highly complex and involve both direct and indirect pathways. Important factors may also include the patient's susceptibility to infection and their innate inflammatory and immune responses.¹⁶⁵

RISK PREDICTION OF PLAQUE INSTABILITY

Imaging

Angiography can demonstrate ulceration¹⁶⁶ but does not appear to be able to adequately distinguish between stable and unstable plaques.¹⁶⁷ In addition, the degree of stenosis detected by angiography does not correlate well with the future risk of events¹⁹⁻²¹ because, as already discussed, it is often not the most stenotic plaques that are at highest risk of rupture.

Conventional ultrasound studies have shown an association between carotid plaque morphology and neurological symptoms¹⁶⁸ but have been unable to predict the risk of future events.23 Intravenous ultrasound (IVUS), however, has been shown to have much greater resolution (100 µm) and provides detailed cross-sectional images of the arterial wall. It is also able to identify the increased echolucence of lipid-rich plaques and for a time it was thought that it might prove useful in detection of rupture-prone plaques.¹⁶⁹ Unfortunately, sensitivity and specificity were found to be low with this technique and it has largely been superseded by intravenous ultrasound virtual histology (IVUS-VH). The improved spectral analysis offered by this technology allows more detailed plaque characterization and can provide detail on lipid content, calcification and volume of the necrotic core.170 Recent studies have shown this may be a clinically useful tool and demonstrated that IVUS-VH identified more TCFAs in patients with acute coronary syndrome than in those with stable angina pectoris.171

In parallel with the development of IVUS-VH, many groups have now begun

to use optical coherence tomography (OCT). Also an intravenous modality, OCT is analogous to ultrasound imaging (using light rather than sound waves) and provides excellent spatial resolution (10-15µm). This allows detailed assessment of the arterial wall and can identify those plaques with a fibrous cap less than 65µm thick (i.e. TCFA) as well as areas of increased echolucency.172 Whilst this technique has yielded very encouraging results in the identification of culprit atherosclerotic lesions, it is not without its limitations. Since blood attenuates the optical signal, the vessel under investigation must be proximally occluded for considerable periods to allow accurate imaging. An updated version of the technology has therefore been developed in recent years. This second generation of OCT is known as optical frequency domain imaging (OFDI) and involves much higher frame rates (>100 frames/sec). The higher frame rate allows rapid three-dimensional imaging of long arterial segments using highspeed pull-back of the probe. This means there is no need for proximal occlusion of the vessel and the artery can simply be purged with saline just prior to imaging¹⁷³ Further investigation will be needed to assess the true clinical utility of this technique.

Since increased inflammatory activity occurs prior to plaque rupture, attempts have been made to detect this increase, using local temperature measurements. Thermography studies have shown that temperature correlates well with macrophage cell density in human carotid plaques.¹⁷⁴ The temperature of coronary vessels in patients with ischaemic heart disease, in particular acute coronary syndromes, is higher than in normal controls.¹⁷⁵ In addition, increased local plaque temperature has been shown to be an independent predictor of adverse clinical outcome.¹⁷⁶

High-resolution MRI appears to characterize the atherosclerotic plaque better than other imaging techniques.¹⁷⁷ It is more accurate then angiography in measuring the degree of stenosis and, unlike angiography and IVUS, is non-invasive. However, a multicentre trial of imaging in coronary artery disease found whilst that MRI could reliably identify significant intraluminal lesions and rule out proximal or three-vessel disease, specificity was low.¹⁷⁸ This led to the suggestion that MRI may be more sensitive and specific if combined with intravascular enhancing agents such as gadolinium. Using such a marker improved MRI specificity and facilitated identification of carotid TCFA.¹⁷⁹

MRI is still technically limited in many cases by small vessel size and movement artefact, and studies have not yet demonstrated the ability to predict risk of future cardiovascular events. Nonetheless, advances in the technique suggest a potential future role for MRI in detection of the high-risk plaque.

Just as enhancing agents may increase the accuracy of MRI, they may also prove useful in identifying atherosclerotic lesions using positron emission tomography (PET) and there has been increasing interest in the use of¹⁸ fluorodeoxyglucose (¹⁸FDG). Uptake of this glucose analogue is increased in metabolically active cells and early animal studies suggest it enriches in plaque macrophages and indicates areas of neovascularisation.¹⁸⁰ However, the clinical application of this technique has yet to be demonstrated.

Blood markers

It has long been established that adverse lipid profiles correlate with increased risk of MI and stroke though this is not a direct predictor of plaque rupture. Raised CRP levels have also been associated with increased cardiovascular risk in apparently healthy patients,^{181,182} though its use as a prognostic marker of clinically significant thrombosis remains controversial. MMP-2 and MMP-9 are raised in the peripheral blood of patients suffering from acute coronary syndromes,¹⁸³ while plasma MMP-9 is raised in patients with unstable carotid plaques.¹⁸⁴ A recent study of 1127 patients with coronary artery disease identified baseline plasma MMP-9 levels to be a novel predictor of cardiovascular mortality.¹⁸⁵

Similarly, raised serum levels of soluble intercellular adhesion molecule-1 (slCAM-1) have been shown to be an independent predictor of future coronary event in patients with coronary heart disease.¹⁸⁶

Many other molecules have also been investigated as potential prognostic markers in progression of atherosclerosis, including cytokines, lipoproteins, myeloperoxidases and placental growth factor. Though some have yielded promising results, none has yet been widely accepted as a reliable predictor of plaque rupture or clinical events.¹⁸⁷

THERAPY AIMED AT PLAQUE STABILISATION

Pharmacotherapy to induce plaque stabilisation could be targeted at different aspects of the complex pathway leading up to plaque rupture, in particular:

- 1. the endothelium by increasing endothelial passivation
- the lipid core by reducing LDL deposition/ augmenting LDL removal
- the fibrous cap by increasing collagen deposition/ preventing collagen degradation
- 4. the vessel lumen by altering the thrombogenicity of the local environment.

Most recent interest has focussed on the role of HMG Co-A reductase inhibitors, which appear capable of influencing plaque stabilisation at all these levels.

HMG Co-A Reductase Inhibitors

HMG Co-A reductase inhibitors, or statins, are well known for their lipid-lowering action. They are the most effective group of therapeutic agents for lowering LDL and raising HDL levels. However, recent evidence suggests that they are also capable of decreasing cardiovascular events in those with normal cholesterol levels.188,189 The Oxford Heart Protection Study¹⁸⁹ was a randomised controlled trial of simvastatin versus placebo in 20,536 individuals at high-risk of cardiovascular disease. Coronary death rate and other vascular events were significantly reduced in the simvastatin groups, even in patients with lipid levels below currently recommended targets (<5mmol/l total cholesterol and <3mmol/l LDL-cholesterol).

In the lipid lowering arm of the Anglo-Scandinavian Cardiac Outcomes Trial (ASCOT),¹⁸⁸ 10,305 individuals with total cholesterol levels <6.5mmol/l were randomised to either atorvastatin or placebo. The trial was stopped 1.7 years before the planned 5-year follow-up target was reached, as there were significantly fewer cardiovascular events in the atorvastatin group. The observed clinical benefit is probably a combination of lipid lowering below levels previously considered 'normal' and additional lipid-independent plaque stabilising actions. Several studies have reported effects other than lipid-lowering properties, including anti-proteolytic and anti-inflammatory mechanisms.^{190,191}

Statins increase nitric oxide synthase activity¹⁹² and encourage endothelial passivation (Figure 4.2). As discussed earlier, nitric oxide causes vasodilatation, inhibition of SMC proliferation and platelet aggregation and has widespread anti-inflammatory and anti-oxidant properties. Statins also reduce the expression of cell adhesion molecules,¹⁹³ interfering with the adherence of monocytes to the endothelium.

Statins may also have direct antiinflammatory and anti-proteolytic actions, which contribute to increased plaque stability. In cell culture and animal models, statins have been shown to reduce macrophage secretion of MMP-1, -2, -3 and -9,¹⁹⁴ and increase the collagen content of the plaque.¹⁹⁵ Also, CRP levels are decreased by statins in a lipidindependent manner.^{191,196}

Work from our laboratories suggests that statin therapy stabilises carotid plaques by lowering the levels of MMP-1, MMP-9 and IL-6. In an observational non-randomised study of 137 patients, we found that patients on statin therapy were significantly less likely to have suffered carotid territory symptoms within the month prior to carotid endarterectomy. The number of patients undergoing spontaneous pre-operative cerebral embolization was also significantly lower in the statin group.

HMG Co-A reductase inhibitors also have the potential to reduce thrombogenicity by decreasing tissue factor activity¹⁹⁷ and lowering levels of PAI-1.^{198,199}

MMP Inhibition

The realisation that tissue remodelling due to increased MMP activity plays a key role in disease states has led to considerable interest in the potential for MMP inhibition. Most clinical and pre-clinical data regarding therapeutic manipulation of the extracellular matrix has been in the fields of arthritis, periodontal disease and cancer.¹⁰³ MMP inhibition aimed at plaque stabilisation aims to redress the imbalance between enzymes and inhibitors, which causes excessive tissue degradation. Potential methods of MMP inhibition include the administration of:

Tissue Inhibitors of Metalloproteinases (TIMPs)

The level of TIMPs can be increased either by the exogenous administration of recombinant TIMPs or by stimulating their local production through gene therapy. Increased TIMP-1 raised the collagen, elastin and smooth muscle content of atherosclerotic lesions in animal models,²⁰⁰ while local gene transfer of TIMP-2 has been shown to decrease vascular remodelling in conjunction with lowered MMP activity (experimental models).²⁰¹

It is difficult to extrapolate these data to potential applications in humans. The major drawback associated with TIMPs would be tissue delivery, since exogenous products would be metabolised and denatured with minimal tissue penetration at the intended site of action. Systemic stimulation of TIMPs would almost certainly have significant side effects precluding clinical use. Therefore, treatment would have to take the form of local tissue delivery or gene therapy. Clearly either system will be very expensive to develop, so more interest has concentrated on the development of synthetic MMP inhibitors.

Synthetic MMP inhibitors

Synthetic peptides work by binding to the zinc ion at the active site of the MMP, thus preventing cleavage of substrate collagen molecules.²⁰² Batimastat showed promise in decreasing tumour development and metastasis (animal models)²⁰³ and limiting aneurysm expansion (experimental models),²⁰⁴ but is not available in an oral form. Marimastat, which is available orally, was shown to limit intimal hyperplasia²⁰⁵ and aneurysm expansion in vivo.²⁰⁶ It also showed promise in early human cancer studies, but caused significant musculoskeletal side effects

in 30% of patients.²⁰⁷ Recent studies of MMI270, a more specific inhibitor (of MMP-2, MMP-8 and MMP-9), have shown a similar side effect profile.²⁰⁸ Furthermore, recent animal studies of broad-spectrum synthetic MMP inhibitors have found them to be generally deleterious in terms of both plaque growth and plaque stability.²⁰⁹

Doxycycline

Doxycycline, a member of the tetracycline antibiotic family, is also a non-selective MMP inhibitor,²¹⁰ with a proven safety profile. Clinical trials have shown that doxycycline is capable of decreasing cartilage MMP levels when given to patients prior to hip surgery.²¹¹ It has also been shown to limit intimal hyperplasia²¹² and aneurysm expansion in vivo,²¹³ by reducing MMP-9 activity. Furthermore, when given to patients prior to AAA repair the expression of MMP-2 and MMP-9 was reduced in the aortic wall.²¹⁴

A randomised clinical trial of doxycycline versus placebo in patients prior to carotid endarterectomy demonstrated decreased plaque MMP-1 levels and a potential for clinical benefit.²¹⁵ A phase II study of doxycycline administration to patients with small AAAs recently showed that it was reasonably well-tolerated (92% completed the 6-month course) and reduced plasma MMP-9 levels.²¹⁶ Further studies are on going to evaluate its effects on small aneurysm expansion.

ACE Inhibitors

ACE inhibitors (ACEI) and angiotensin II receptor antagonists decrease cardiovascular events, independently of their effects on blood pressure control. The ACEI, trandalopril, and the experimental angiotensin receptor antagonist, HR720, decrease the area

of atherosclerotic lesions in the thoracic aorta of cholesterol-fed monkeys.²¹⁷ This was achieved without alteration of mean blood pressure or cholesterol levels. The Heart Outcomes Prevention Evaluation (HOPE) study demonstrated a decrease in cardiovascular events in high-risk patients given ramipril as opposed to placebo.²¹⁸ This effect could only be partly explained by the modest decrease in mean blood pressure seen between the 2 groups (3/2mmHg).

Angiotensin II promotes endothelial activation,²¹⁹ and therefore the mechanism of action of ACEIs could be through endothelial passivation (leading to a reduction in cell adhesion molecule expression and macrophage infiltration). ACEIs may also exert their effects through bradykinin potentiation, resulting in decreased smooth muscle cell migration, decreased inflammation and decreased production of oxygen free radicals (COPPOLA 2008). Navalkar et al provided biochemical evidence to support these hypotheses by demonstrating that irbesartan (an angiotensin II receptor blocker) can decrease plasma levels of VCAM-1, TNF- α and superoxide.220

With ever more detailed understanding of the human genome, gene therapies have also come under investigation in the search for anti-atherosclerotic therapies. Hans et al have demonstrated that polyADP-ribose polymerase (PARP-1), a DNA-repair protein, stimulates apoptosis in the presence of local inflammation and plays an important role in plaque dynamics. They went on to show that inhibition of PARP-1 resulted in a reduction in plaque size, decreased collagen degradation and increased plaque smooth muscle content in ApoE(-/-) mice.²²¹ These findings suggest that PARP-1 inhibition may also represent a valuable therapeutic tool, though its applicability in humans has yet to be demonstrated.

Since the clinical significance of any plaque

rupture is also governed by the intravascular environment, investigators continue to seek new therapies that may decrease the thrombogenicity of blood. Until now, this has been achieved with a combination of aspirin and another antiplatelet agent most commonly clopidogrel - but drug resistance and side effect profiles can limit its applicability. The latest class of antiplatelet drugs is the P2Y₁₂ blockers, which inhibits platelet activation via blockade of the P2Y₁₂ ADP-receptor. Though these drugs (such as prasugrel and ticagrelor) may still have significant side effect profiles, they seem to be associated with far less unwanted bleeding and may be effective in patients who do not respond to clopidogrel.²²²

Though a number of therapeutic targets have shown promise in preventing plaque rupture, substantial work is still needed in this area since many of the potential therapeutic targets (such as smooth muscle cells and macrophages) have the ability to play both detrimental and beneficial roles in the complex process of atherosclerosis.

SUMMARY

Acute plaque disruption precedes the onset of clinical ischaemic syndromes. Exposure of the highly thrombogenic core to luminal blood results in platelet adherence and thrombosis. Inflammation is clearly involved in the process of plaque development and acute disruption, though the precise mechanism by which the inflammatory process is initiated remains unclear. The roles of angiogenesis, cellular apoptosis and infectious agents also require further clarification. Unstable plaques have a large lipid core and a thin fibrous cap with reduced collagen content. A major component of plaque destabilisation appears to be increased matrix degradation, the primary regulators of which are the MMPs and their inhibitors. There are a number

of potential therapeutic options aimed at preventing plaque disruption. In particular, MMP inhibition is an attractive target for such pharmacotherapy.

REFERENCES

- Casscells W, Naghavi M, Willerson JT. Vulnerable atherosclerotic plaque: a multifocal disease. *Circulation* 2003; 107(16): 2072–5.
- Little WC. Angiographic assessment of the culprit coronary artery lesion before acute myocardial infarction. *The American journal of cardiology* 1990; **66**(16): 44G–47G.
- Naghavi M, Libby P, Falk E, Casscells SW, Litovsky S, Rumberger J, et al. From vulnerable plaque to vulnerable patient: a call for new definitions and risk assessment strategies: Part I. *Circulation* 2003; 108(14): 1664–72.
- Falk E, Shah PK, Fuster V. Coronary plaque disruption. *Circulation* 1995; 92(3): 657–71.
- Libby P. Current concepts of the pathogenesis of the acute coronary syndromes. *Circulation* 2001; **104**(3): 365–72.
- Shah PK. Plaque disruption and thrombosis: potential role of inflammation and infection. *Cardiology in review* 2000; 8(1): 31–9.
- Falk E. Unstable angina with fatal outcome: dynamic coronary thrombosis leading to infarction and/or sudden death. Autopsy evidence of recurrent mural thrombosis with peripheral embolization culminating in total vascular occlusion. *Circulation* 1985; **71**(4): 699–708.
- 8. Carr S, Farb A, Pearce WH, Virmani R, Yao JS. Atherosclerotic plaque rupture in symptomatic carotid artery stenosis.

J Vasc Surg 1996; **23**(5): 755–65; discussion 65–6.

- Loftus IM, Naylor AR, Goodall S, Crowther M, Jones L, Bell PR, et al. Increased matrix metalloproteinase-9 activity in unstable carotid plaques. A potential role in acute plaque disruption. *Stroke* 2000; **31**(1): 40–7.
- Sitzer M, Müller W, Siebler M, Hort W, Kniemeyer HW, Jäncke L, et al. Plaque ulceration and lumen thrombus are the main sources of cerebral microemboli in high-grade internal carotid artery stenosis. *Stroke* 1995; **26**(7): 1231–3.
- Davies MJ, Thomas A. Thrombosis and acute coronary-artery lesions in sudden cardiac ischemic death. *N Engl* J Med 1984; **310**(18): 1137–40.
- Falk E. Plaque rupture with severe preexisting stenosis precipitating coronary thrombosis. Characteristics of coronary atherosclerotic plaques underlying fatal occlusive thrombi. *Br Heart J* 1983; 50(2): 127–34.
- Friedman M. Pathogenesis of coronary thrombosis, intramural and intraluminal hemorrhage. *Advances in cardiology* 1970; 4: 20–46.
- Forrester JS, Litvack F, Grundfest W, Hickey A. A perspective of coronary disease seen through the arteries of living man. *Circulation* 1987; 75(3): 505–13.
- Sherman CT, Litvack F, Grundfest W, Lee M, Hickey A, Chaux A, et al. Coronary angioscopy in patients with unstable angina pectoris. *N Engl J Med* 1986; **315**(15): 913–9.
- Davies MJ. The pathophysiology of acute coronary syndromes. *Heart* 2000; 83(3): 361–6.
- 17. Felton CV, Crook D, Davies MJ, Oliver MF. Relation of plaque lipid composition and morphology to the stability of human aortic plaques.

Arterioscler Thromb Vasc Biol 1997; **17**(7): 1337–45.

- Kolodgie FD, Burke AP, Farb A, Gold HK, Yuan J, Narula J, et al. The thin-cap fibroatheroma: a type of vulnerable plaque: the major precursor lesion to acute coronary syndromes. *Curr Opin Cardiol* 2001; 16(5): 285–92.
- Ambrose JA, Winters SL, Stern A, Eng A, Teichholz LE, Gorlin R, et al. Angiographic morphology and the pathogenesis of unstable angina pectoris. *Journal of the American College* of Cardiology 1985; 5(3): 609–16.
- Giroud D, Li JM, Urban P, Meier B, Rutishauer W. Relation of the site of acute myocardial infarction to the most severe coronary arterial stenosis at prior angiography. *The American journal of cardiology* 1992; **69**(8): 729–32.
- Hackett D, Davies G, Maseri A. Preexisting coronary stenoses in patients with first myocardial infarction are not necessarily severe. *European Heart Journal* 1988; 9(12): 1317–23.
- 22. Markus HS, Thomson ND, Brown MM. Asymptomatic cerebral embolic signals in symptomatic and asymptomatic carotid artery disease. *Brain* 1995; **118** (Pt 4): 1005–11.
- Golledge J, Cuming R, Ellis M, Davies AH, Greenhalgh RM. Carotid plaque characteristics and presenting symptom. *Br J Surg* 1997; 84(12): 1697–701.
- Gunning A, Pickering G, Robb-Smith A, Russell R. Mural thrombosis of the subclavian artery and subsequent embolism in cervical rib Q J Med 1964; 33: 133–54.
- 25. Harrison MJ, Marshall J. The finding of thrombus at carotid endarterectomy and its relationship to the timing of surgery. *Br J Surg* 1977; **64**(7): 511–2.

- 26. Bogousslavsky J, Van Melle G, Regli F. The Lausanne Stroke Registry: analysis of 1,000 consecutive patients with first stroke. *Stroke* 1988; **19**(9): 1083–92.
- 27. Imparato AM, Riles TS, Gorstein F. The carotid bifurcation plaque: pathologic findings associated with cerebral ischemia. *Stroke* 1979; **10**(3): 238–45.
- Persson AV, Robichaux WT, Silverman M. The natural history of carotid plaque development. *Arch Surg* 1983; 118(9): 1048–52.
- Lusby RJ, Ferrell LD, Ehrenfeld WK, Stoney RJ, Wylie EJ. Carotid plaque hemorrhage. Its role in production of cerebral ischemia. *Arch Surg* 1982; 117(11): 1479–88.
- Tenaglia AN, Peters KG, Sketch MH, Annex BH. Neovascularization in atherectomy specimens from patients with unstable angina: implications for pathogenesis of unstable angina. *American heart journal* 1998; 135(1): 10–4.
- Van Damme H, Vivario M. Pathologic aspects of carotid plaques: surgical and clinical significance. *Int Angiol* 1993; 12(4): 299–311.
- 32. Barua RS, Ambrose JA, Srivastava S, DeVoe MC, Eales-Reynolds L-J. Reactive oxygen species are involved in smoking-induced dysfunction of nitric oxide biosynthesis and upregulation of endothelial nitric oxide synthase: an in vitro demonstration in human coronary artery endothelial cells. *Circulation* 2003; **107**(18): 2342–7.
- Barua RS, Ambrose JA, Saha DC, Eales-Reynolds L-J. Smoking is associated with altered endothelialderived fibrinolytic and antithrombotic factors: an in vitro demonstration. *Circulation* 2002; 106(8): 905–8.

- 34. Hanratty CG, McGrath LT, McAuley DF, Young IS, Johnston GD. The effects of oral methionine and homocysteine on endothelial function. *Heart* 2001; 85(3): 326–30.
- Lüscher TF, Tanner FC, Noll G. Lipids and endothelial function: effects of lipid-lowering and other therapeutic interventions. *Current opinion in lipidology* 1996; 7(4): 234–40.
- 36. Salt IP, Morrow VA, Brandie FM, Connell JMC, Petrie JR. High glucose inhibits insulin-stimulated nitric oxide production without reducing endothelial nitric-oxide synthase Ser1177 phosphorylation in human aortic endothelial cells. *J Biol Chem* 2003; **278**(21): 18791–7.
- Taddei S, Virdis A, Ghiadoni L, Sudano I, Salvetti A. Endothelial dysfunction in hypertension. *J Cardiovasc Pharmacol* 2001; 38 Suppl 2: S11–4.
- Cybulsky MI, Gimbrone MA. Endothelial expression of a mononuclear leukocyte adhesion molecule during atherogenesis. *Science* 1991; **251**(4995): 788–91.
- 39. van der Wal AC, Das PK, Tigges AJ, Becker AE. Adhesion molecules on the endothelium and mononuclear cells in human atherosclerotic lesions. *Am J Pathol* 1992; 141(6): 1427–33.
- Vanhoutte PM. Endothelial dysfunction and atherosclerosis. *European Heart Journal* 1997; 18 Suppl E: E19–29.
- 41. De Caterina R, Libby P, Peng HB, Thannickal VJ, Rajavashisth TB, Gimbrone MA, et al. Nitric oxide decreases cytokine-induced endothelial activation. Nitric oxide selectively reduces endothelial expression of adhesion molecules and proinflammatory cytokines. *J Clin Invest* 1995; **96**(1): 60–8.

- Moncada S, Higgs A. The L-argininenitric oxide pathway. N Engl J Med 1993; 329(27): 2002–12.
- Kingwell BA, Sherrard B, Jennings GL, Dart AM. Four weeks of cycle training increases basal production of nitric oxide from the forearm. *Am J Physiol* 1997; 272(3 Pt 2): H1070–7.
- Malek AM, Alper SL, Izumo S. Hemodynamic shear stress and its role in atherosclerosis. *Jama* 1999; 282(21): 2035–42.
- Gimbrone MA, Resnick N, Nagel T, Khachigian LM, Collins T, Topper JN. Hemodynamics, endothelial gene expression, and atherogenesis. *Ann NY Acad Sci* 1997; **811**: 1–10; discussion 10–1.
- 46. Nadaud S, Philippe M, Arnal JF, Michel JB, Soubrier F. Sustained increase in aortic endothelial nitric oxide synthase expression in vivo in a model of chronic high blood flow. *Circ Res* 1996; **79**(4): 857–63.
- De Keulenaer GW, Chappell DC, Ishizaka N, Nerem RM, Alexander RW, Griendling KK. Oscillatory and steady laminar shear stress differentially affect human endothelial redox state: role of a superoxide-producing NADH oxidase. *Circ Res* 1998; **82**(10): 1094–101.
- Chappell DC, Varner SE, Nerem RM, Medford RM, Alexander RW. Oscillatory shear stress stimulates adhesion molecule expression in cultured human endothelium. *Circ Res* 1998; 82(5): 532–9.
- Lee RT, Kamm RD. Vascular mechanics for the cardiologist. *Journal* of the American College of Cardiology 1994; 23(6): 1289–95.
- 50. Cheng GC, Loree HM, Kamm RD, Fishbein MC, Lee RT. Distribution of circumferential stress in

ruptured and stable atherosclerotic lesions. A structural analysis with histopathological correlation. *Circulation* 1993; **87**(4): 1179–87.

- Loree HM, Kamm RD, Stringfellow RG, Lee RT. Effects of fibrous cap thickness on peak circumferential stress in model atherosclerotic vessels. *Circ Res* 1992; 71(4): 850–8.
- 52. MacIsaac AI, Thomas JD, Topol EJ. Toward the quiescent coronary plaque. *Journal of the American College of Cardiology* 1993; **22**(4): 1228–41.
- Grønholdt ML, Dalager-Pedersen S, Falk E. Coronary atherosclerosis: determinants of plaque rupture. *European Heart Journal* 1998; 19 Suppl C: C24–9.
- 54. Kragel AH, Reddy SG, Wittes JT, Roberts WC. Morphometric analysis of the composition of atherosclerotic plaques in the four major epicardial coronary arteries in acute myocardial infarction and in sudden coronary death. *Circulation* 1989; **80**(6): 1747–56.
- 55. Gertz SD, Roberts WC. Hemodynamic shear force in rupture of coronary arterial atherosclerotic plaques. *The American journal of cardiology* 1990; **66**(19): 1368–72.
- Davies MJ, Richardson PD, Woolf N, Katz DR, Mann J. Risk of thrombosis in human atherosclerotic plaques: role of extracellular lipid, macrophage, and smooth muscle cell content. *Br Heart J* 1993; 69(5): 377–81.
- 57. el-Barghouti N, Nicolaides AN, Tegos T, Geroulakos G. The relative effect of carotid plaque heterogeneity and echogenicity on ipsilateral cerebral infarction and symptoms of cerebrovascular disease. *Int Angiol* 1996; **15**(4): 300–6.

- Reilly LM, Lusby RJ, Hughes L, Ferrell LD, Stoney RJ, Ehrenfeld WK. Carotid plaque histology using realtime ultrasonography. Clinical and therapeutic implications. *Am J Surg* 1983; 146(2): 188–93.
- Feeley TM, Leen EJ, Colgan MP, Moore DJ, Hourihane DO, Shanik GD. Histologic characteristics of carotid artery plaque. *J Vasc Surg* 1991; 13(5): 719–24.
- 60. Grønholdt M-LM, Nordestgaard BG, Bentzon J, Wiebe BM, Zhou J, Falk E, et al. Macrophages are associated with lipid-rich carotid artery plaques, echolucency on B-mode imaging, and elevated plasma lipid levels. *J Vasc Surg* 2002; **35**(1): 137–45.
- Takano M, Mizuno K, Okamatsu K, Yokoyama S, Ohba T, Sakai S. Mechanical and structural characteristics of vulnerable plaques: analysis by coronary angioscopy and intravascular ultrasound. *Journal of the American College of Cardiology* 2001; 38(1): 99–104.
- Steinberg D. Low density lipoprotein oxidation and its pathobiological significance. *J Biol Chem* 1997; 272(34): 20963–6.
- Nicholson AC, Han J, Febbraio M, Silversterin RL, Hajjar DP. Role of CD36, the macrophage class B scavenger receptor, in atherosclerosis. *Ann N Y Acad Sci* 2001; 947: 224–8.
- Kruth HS, Huang W, Ishii I, Zhang W-Y. Macrophage foam cell formation with native low density lipoprotein. *J Biol Chem* 2002; 277(37): 34573–80.
- 65. de la Llera Moya M, Atger V, Paul JL, Fournier N, Moatti N, Giral P, et al. A cell culture system for screening human serum for ability to promote cellular cholesterol efflux. Relations

between serum components and efflux, esterification, and transfer. *Arterioscler Thromb* 1994; **14**(7): 1056–65.

- 66. Badimon JJ, Badimon L, Fuster V. Regression of atherosclerotic lesions by high density lipoprotein plasma fraction in the cholesterol-fed rabbit. *J Clin Invest* 1990; **85**(4): 1234–41.
- Spieker LE, Sudano I, Hürlimann D, Lerch PG, Lang MG, Binggeli C, et al. High-density lipoprotein restores endothelial function in hypercholesterolemic men. *Circulation* 2002; **105**(12): 1399–402.
- Barter PJ. Inhibition of endothelial cell adhesion molecule expression by high density lipoproteins. *Clin Exp Pharmacol Physiol* 1997; 24(3–4): 286–7.
- Lin K-Y, Chen Y-L, Shih C-C, Pan J-P, Chan W-E, Chiang A-N. Contribution of HDL-apolipoproteins to the inhibition of low density lipoprotein oxidation and lipid accumulation in macrophages. *J Cell Biochem* 2002; 86(2): 258–67.
- Abela GS, Aziz K. Cholesterol crystals rupture biological membranes and human plaques during acute cardiovascular events--a novel insight into plaque rupture by scanning electron microscopy. *Scanning* 2006; 28(1): 1–10.
- Vedre A, Pathak DR, Crimp M, Lum C, Koochesfahani M, Abela GS. Physical factors that trigger cholesterol crystallization leading to plaque rupture. *Atherosclerosis* 2009; 203(1): 89–96.
- 72. Koskinas KC, Chatzizisis YS, Baker AB, Edelman ER, Stone PH, Feldman CL. The role of low endothelial shear stress in the conversion of atherosclerotic lesions from stable to unstable plaque. *Curr Opin Cardiol* 2009; **24**(6): 580–90.

- 73. Farb A, Burke AP, Tang AL, Liang TY, Mannan P, Smialek J, et al. Coronary plaque erosion without rupture into a lipid core. A frequent cause of coronary thrombosis in sudden coronary death. *Circulation* 1996; **93**(7): 1354–63.
- 74. van der Wal AC, Becker AE, van der Loos CM, Das PK. Site of intimal rupture or erosion of thrombosed coronary atherosclerotic plaques is characterized by an inflammatory process irrespective of the dominant plaque morphology. *Circulation* 1994; **89**(1): 36–44.
- 75. Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM. Lessons from sudden coronary death: a comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol* 2000; **20**(5): 1262–75.
- 76. Burke AP, Kolodgie FD, Farb A, Weber DK, Malcom GT, Smialek J, et al. Healed plaque ruptures and sudden coronary death: evidence that subclinical rupture has a role in plaque progression. *Circulation* 2001; **103**(7): 934–40.
- 77. Burleigh MC, Briggs AD, Lendon CL, Davies MJ, Born GV, Richardson PD. Collagen types I and III, collagen content, GAGs and mechanical strength of human atherosclerotic plaque caps: span-wise variations. *Atherosclerosis* 1992; **96**(1): 71–81.
- Davies MJ. Stability and instability: two faces of coronary atherosclerosis. The Paul Dudley White Lecture 1995. *Circulation* 1996; **94**(8): 2013–20.
- Kockx MM. Apoptosis in the atherosclerotic plaque: quantitative and qualitative aspects. *Arterioscler Thromb Vasc Biol* 1998; 18(10): 1519–22.
- 80. Kockx MM, De Meyer GR, Muhring J, Jacob W, Bult H, Herman AG.

Apoptosis and related proteins in different stages of human atherosclerotic plaques. *Circulation* 1998; **97**(23): 2307–15.

- Vengrenyuk Y, Carlier S, Xanthos S, Cardoso L, Ganatos P, Virmani R, et al. A hypothesis for vulnerable plaque rupture due to stressinduced debonding around cellular microcalcifications in thin fibrous caps. *Proc Natl Acad Sci USA* 2006; **103**(40): 14678–83.
- Barnes MJ, Farndale RW. Collagens and atherosclerosis. *Exp Gerontol* 1999; 34(4): 513–25.
- Dilley RJ, McGeachie JK, Prendergast FJ. A review of the proliferative behaviour, morphology and phenotypes of vascular smooth muscle. *Atherosclerosis* 1987; 63(2–3): 99–107.
- 84. Newby AC. Molecular and cell biology of native coronary and vein-graft atherosclerosis: regulation of plaque stability and vessel-wall remodelling by growth factors and cell-extracellular matrix interactions. *Coron Artery Dis* 1997; 8(3–4): 213–24.
- 85. Amento EP, Ehsani N, Palmer H, Libby P. Cytokines and growth factors positively and negatively regulate interstitial collagen gene expression in human vascular smooth muscle cells. *Arterioscler Thromb* 1991; **11**(5): 1223–30.
- Bentzon JF, Falk E. Circulating smooth muscle progenitor cells in atherosclerosis and plaque rupture: current perspective and methods of analysis. *Vascul Pharmacol* 2010; 52(1–2): 11–20.
- 87. Clarke MCH, Littlewood TD, Figg N, Maguire JJ, Davenport AP, Goddard M, et al. Chronic apoptosis of vascular smooth muscle cells accelerates

atherosclerosis and promotes calcification and medial degeneration. *Circ Res* 2008; **102**(12): 1529–38.

- Geng YJ. Biologic effect and molecular regulation of vascular apoptosis in atherosclerosis. *Curr Atheroscler Rep* 2001; 3(3): 234–42.
- Geng YJ, Libby P. Evidence for apoptosis in advanced human atheroma. Colocalization with interleukin-1 beta-converting enzyme. *Am J Pathol* 1995; 147(2): 251–66.
- Bauriedel G, Hutter R, Welsch U, Bach R, Sievert H, Lüderitz B. Role of smooth muscle cell death in advanced coronary primary lesions: implications for plaque instability. *Cardiovasc Res* 1999; **41**(2): 480–8.
- Bennett MR, Evan GI, Schwartz SM. Apoptosis of human vascular smooth muscle cells derived from normal vessels and coronary atherosclerotic plaques. *J Clin Invest* 1995; **95**(5): 2266–74.
- Buja LM, Willerson JT. Role of inflammation in coronary plaque disruption. *Circulation* 1994; 89(1): 503–5.
- 93. Libby P, Simon DI. Inflammation and thrombosis: the clot thickens. *Circulation* 2001; **103**(13): 1718–20.
- 94. Moreno PR, Falk E, Palacios IF, Newell JB, Fuster V, Fallon JT. Macrophage infiltration in acute coronary syndromes. Implications for plaque rupture. *Circulation* 1994; **90**(2): 775–8.
- 95. Shah PK, Falk E, Badimon JJ, Fernandez-Ortiz A, Mailhac A, Villareal-Levy G, et al. Human monocyte-derived macrophages induce collagen breakdown in fibrous caps of atherosclerotic plaques. Potential role of matrix-degrading metalloproteinases

and implications for plaque rupture. *Circulation* 1995; **92**(6): 1565–9.

- Loftus IM, Naylor AR, Bell PRF, Thompson MM. Matrix metalloproteinases and atherosclerotic plaque instability. *Br J Surg* 2002; 89(6): 680–94.
- 97. Agren MS, Jorgensen LN, Andersen M, Viljanto J, Gottrup F. Matrix metalloproteinase 9 level predicts optimal collagen deposition during early wound repair in humans. *Br J Surg* 1998; **85**(1): 68–71.
- Wysocki AB, Staiano-Coico L, Grinnell F. Wound fluid from chronic leg ulcers contains elevated levels of metalloproteinases MMP-2 and MMP-9. *J Invest Dermatol* 1993; 101(1): 64–8.
- Krane SM. Clinical importance of metalloproteinases and their inhibitors. *Ann N Y Acad Sci* 1994; 732: 1–10.
- 100. Page RC. The role of inflammatory mediators in the pathogenesis of periodontal disease. *J Periodont Res* 1991; **26**(3 Pt 2): 230–42.
- 101. Harris ED. Rheumatoid arthritis. Pathophysiology and implications for therapy. N Engl J Med 1990; **322**(18): 1277–89.
- 102. Parsons SL, Watson SA, Brown PD, Collins HM, Steele RJ. Matrix metalloproteinases. *Br J Surg* 1997; 84(2): 160–6.
- 103. Dollery CM, McEwan JR, Henney AM. Matrix metalloproteinases and cardiovascular disease. *Circ Res* 1995; 77(5): 863–8.
- 104. Mauviel A. Cytokine regulation of metalloproteinase gene expression. *J Cell Biochem* 1993; **53**(4): 288–95.
- 105. Rajavashisth TB, Xu XP, Jovinge S, Meisel S, Xu XO, Chai NN, et al. Membrane type 1 matrix metalloproteinase expression in human

atherosclerotic plaques: evidence for activation by proinflammatory mediators. *Circulation* 1999; **99**(24): 3103–9.

- 106. Singer CF, Marbaix E, Lemoine P, Courtoy PJ, Eeckhout Y. Local cytokines induce differential expression of matrix metalloproteinases but not their tissue inhibitors in human endometrial fibroblasts. *Eur J Biochem* 1999; 259(1–2): 40–5.
- 107. Chen H, Li D, Saldeen T, Mehta JL. TGF-beta 1 attenuates myocardial ischemia-reperfusion injury via inhibition of upregulation of MMP-1. *Am J Physiol Heart Circ Physiol* 2003; 284(5): H1612–7.
- 108. Gogly B, Hornebeck W, Groult N, Godeau G, Pellat B. Influence of heparin(s) on the interleukin-1-betainduced expression of collagenase, stromelysin-1, and tissue inhibitor of metalloproteinase-1 in human gingival fibroblasts. *Biochem Pharmacol* 1998; 56(11): 1447–54.
- 109. Biswas C, Zhang Y, DeCastro R, Guo H, Nakamura T, Kataoka H, et al. The human tumor cell-derived collagenase stimulatory factor (renamed EMMPRIN) is a member of the immunoglobulin superfamily. *Cancer Res* 1995; **55**(2): 434–9.
- 110. Schmidt R, Bültmann A, Fischel S, Gillitzer A, Cullen P, Walch A, et al. Extracellular matrix metalloproteinase inducer (CD147) is a novel receptor on platelets, activates platelets, and augments nuclear factor kappaBdependent inflammation in monocytes. *Circ Res* 2008; **102**(3): 302–9.
- Nagase H. Activation mechanisms of matrix metalloproteinases. *Biol Chem* 1997; **378**(3–4): 151–60.

- Lijnen HR. Plasmin and matrix metalloproteinases in vascular remodeling. *Thromb Haemost* 2001; 86(1): 324–33.
- 113. Galis ZS, Kranzhöfer R, Fenton JW, Libby P. Thrombin promotes activation of matrix metalloproteinase-2 produced by cultured vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 1997; 17(3): 483–9.
- 114. Rajagopalan S, Meng XP, Ramasamy S, Harrison DG, Galis ZS. Reactive oxygen species produced by macrophage-derived foam cells regulate the activity of vascular matrix metalloproteinases in vitro. Implications for atherosclerotic plaque stability. J Clin Invest 1996; **98**(11): 2572–9.
- 115. Xu XP, Meisel SR, Ong JM, Kaul S, Cercek B, Rajavashisth TB, et al. Oxidized low-density lipoprotein regulates matrix metalloproteinase-9 and its tissue inhibitor in human monocyte-derived macrophages. *Circulation* 1999; **99**(8): 993–8.
- 116. Gomez DE, Alonso DF, Yoshiji H, Thorgeirsson UP. Tissue inhibitors of metalloproteinases: structure, regulation and biological functions. *Eur J Cell Biol* 1997; 74(2): 111–22.
- 117. Vine N, Powell JT. Metalloproteinases in degenerative aortic disease. *Clin Sci* 1991; **81**(2): 233–9.
- 118. Thompson RW, Holmes DR, Mertens RA, Liao S, Botney MD, Mecham RP, et al. Production and localization of 92-kilodalton gelatinase in abdominal aortic aneurysms. An elastolytic metalloproteinase expressed by aneurysm-infiltrating macrophages. *J Clin Invest* 1995; **96**(1): 318–26.
- 119. Li Z, Li L, Zielke HR, Cheng L, Xiao R, Crow MT, et al. Increased

expression of 72-kd type IV collagenase (MMP-2) in human aortic atherosclerotic lesions. *Am J Pathol* 1996; **148**(1): 121–8.

- 120. Galis ZS, Sukhova GK, Lark MW, Libby P. Increased expression of matrix metalloproteinases and matrix degrading activity in vulnerable regions of human atherosclerotic plaques. J Clin Invest 1994; 94(6): 2493–503.
- 121. Brown DL, Hibbs MS, Kearney M, Loushin C, Isner JM. Identification of 92-kD gelatinase in human coronary atherosclerotic lesions. Association of active enzyme synthesis with unstable angina. *Circulation* 1995; **91**(8): 2125–31.
- 122. Aimes RT, Quigley JP. Matrix metalloproteinase-2 is an interstitial collagenase. Inhibitor-free enzyme catalyzes the cleavage of collagen fibrils and soluble native type I collagen generating the specific 3/4- and 1/4-length fragments. J Biol Chem 1995; **270**(11): 5872–6.
- 123. Sukhova GK, Schönbeck U, Rabkin E, Schoen FJ, Poole AR, Billinghurst RC, et al. Evidence for increased collagenolysis by interstitial collagenases-1 and -3 in vulnerable human atheromatous plaques. *Circulation* 1999; **99**(19): 2503–9.
- 124. Herman MP, Sukhova GK, Libby P, Gerdes N, Tang N, Horton DB, et al. Expression of neutrophil collagenase (matrix metalloproteinase-8) in human atheroma: a novel collagenolytic pathway suggested by transcriptional profiling. *Circulation* 2001; **104**(16): 1899–904.
- 125. Lee RT, Schoen FJ, Loree HM, Lark MW, Libby P. Circumferential stress and matrix metalloproteinase 1 in human coronary atherosclerosis.

Implications for plaque rupture. *Arterioscler Thromb Vasc Biol* 1996; **16**(8): 1070–3.

- 126. Arroyo LH, Lee RT. Mechanisms of plaque rupture: mechanical and biologic interactions. *Cardiovasc Res* 1999; **41**(2): 369–75.
- 127. Price SJ, Greaves DR, Watkins H. Identification of novel, functional genetic variants in the human matrix metalloproteinase-2 gene: role of Sp1 in allele-specific transcriptional regulation. *J Biol Chem* 2001; 276(10): 7549–58.
- 128. Ye S, Watts GF, Mandalia S, Humphries SE, Henney AM. Preliminary report: genetic variation in the human stromelysin promoter is associated with progression of coronary atherosclerosis. *Br Heart J* 1995; **73**(3): 209–15.
- 129. Terashima M, Akita H, Kanazawa K, Inoue N, Yamada S, Ito K, et al. Stromelysin promoter 5A/6A polymorphism is associated with acute myocardial infarction. *Circulation* 1999; **99**(21): 2717–9.
- 130. Zhang B, Henney A, Eriksson P, Hamsten A, Watkins H, Ye S. Genetic variation at the matrix metalloproteinase-9 locus on chromosome 20q12.2-13.1. *Hum Genet* 1999; **105**(5): 418–23.
- 131. Nemerson Y. Tissue factor and hemostasis. *Blood* 1988; 71(1): 1–8.
- 132. Fernández-Ortiz A, Badimon JJ, Falk E, Fuster V, Meyer B, Mailhac A, et al. Characterization of the relative thrombogenicity of atherosclerotic plaque components: implications for consequences of plaque rupture. *Journal of the American College of Cardiology* 1994; **23**(7): 1562–9.
- 133. Moreno PR, Bernardi VH, López-Cuéllar J, Murcia AM,

Palacios IF, Gold HK, et al. Macrophages, smooth muscle cells, and tissue factor in unstable angina. Implications for cell-mediated thrombogenicity in acute coronary syndromes. *Circulation* 1996; **94**(12): 3090–7.

- Mach F, Schönbeck U, Libby P. CD40 signaling in vascular cells: a key role in atherosclerosis? *Atherosclerosis* 1998; 137 Suppl: S89–95.
- 135. Aikawa M, Voglic SJ, Sugiyama S, Rabkin E, Taubman MB, Fallon JT, et al. Dietary lipid lowering reduces tissue factor expression in rabbit atheroma. *Circulation* 1999; **100**(11): 1215–22.
- 136. Bevilacqua MP, Schleef RR, Gimbrone MA, Loskutoff DJ. Regulation of the fibrinolytic system of cultured human vascular endothelium by interleukin 1. *J Clin Invest* 1986; **78**(2): 587–91.
- 137. Giesen PL, Rauch U, Bohrmann B, Kling D, Roqué M, Fallon JT, et al. Blood-borne tissue factor: another view of thrombosis. *Proc Natl Acad Sci* USA 1999; **96**(5): 2311–5.
- 138. Meisel SR, Xu XP, Edgington TS, Dimayuga P, Kaul S, Lee S, et al. Differentiation of adherent human monocytes into macrophages markedly enhances tissue factor protein expression and procoagulant activity. *Atherosclerosis* 2002; **161**(1): 35–43.
- Toschi V, Gallo R, Lettino M, Fallon JT, Gertz SD, Fernández-Ortiz A, et al. Tissue factor modulates the thrombogenicity of human atherosclerotic plaques. *Circulation* 1997; **95**(3): 594–9.
- 140. Matetzky S, Tani S, Kangavari S, Dimayuga P, Yano J, Xu H, et al. Smoking increases tissue factor

expression in atherosclerotic plaques: implications for plaque thrombogenicity. *Circulation* 2000; **102**(6): 602–4.

- 141. Sambola A, Osende J, Hathcock J, Degen M, Nemerson Y, Fuster V, et al. Role of risk factors in the modulation of tissue factor activity and blood thrombogenicity. *Circulation* 2003; 107(7): 973–7.
- 142. Barger AC, Beeuwkes R, Lainey LL, Silverman KJ. Hypothesis: vasa vasorum and neovascularization of human coronary arteries. A possible role in the pathophysiology of atherosclerosis. *N Engl J Med* 1984; **310**(3): 175–7.
- 143. Virmani R, Kolodgie FD, Burke AP, Finn AV, Gold HK, Tulenko TN, et al. Atherosclerotic plaque progression and vulnerability to rupture: angiogenesis as a source of intraplaque hemorrhage. *Arterioscler Thromb Vasc Biol* 2005; **25**(10): 2054–61.
- 144. de Boer OJ, van der Wal AC, Teeling P, Becker AE. Leucocyte recruitment in rupture prone regions of lipidrich plaques: a prominent role for neovascularization? *Cardiovasc Res* 1999; **41**(2): 443–9.
- 145. Haas TL, Madri JA. Extracellular matrix-driven matrix metalloproteinase production in endothelial cells: implications for angiogenesis. *Trends Cardiovasc Med* 1999; 9(3–4): 70–7.
- 146. Kostoulas G, Lang A, Nagase H, Baici A. Stimulation of angiogenesis through cathepsin B inactivation of the tissue inhibitors of matrix metalloproteinases. *FEBS Lett* 1999; 455(3): 286–90.
- 147. Muhlestein JB, Anderson JL, Hammond EH, Zhao L, Trehan S, Schwobe EP, et al. Infection with

Chlamydia pneumoniae accelerates the development of atherosclerosis and treatment with azithromycin prevents it in a rabbit model. *Circulation* 1998; **97**(7): 633–6.

- 148. Saikku P, Leinonen M, Tenkanen L, Linnanmäki E, Ekman MR, Manninen V, et al. Chronic Chlamydia pneumoniae infection as a risk factor for coronary heart disease in the Helsinki Heart Study. Ann Intern Med 1992; 116(4): 273–8.
- 149. Saikku P, Leinonen M, Mattila K, Ekman MR, Nieminen MS, Mäkelä PH, et al. Serological evidence of an association of a novel Chlamydia, TWAR, with chronic coronary heart disease and acute myocardial infarction. *Lancet* 1988; 2(8618): 983–6.
- 150. Ossei-Gerning N, Moayyedi P, Smith S, Braunholtz D, Wilson JI, Axon AT, et al. Helicobacter pylori infection is related to atheroma in patients undergoing coronary angiography. *Cardiovasc Res* 1997; **35**(1): 120–4.
- 151. Nieto FJ, Adam E, Sorlie P, Farzadegan H, Melnick JL, Comstock GW, et al. Cohort study of cytomegalovirus infection as a risk factor for carotid intimal-medial thickening, a measure of subclinical atherosclerosis. *Circulation* 1996; 94(5): 922–7.
- 152. Span AH, Grauls G, Bosman F, van Boven CP, Bruggeman CA. Cytomegalovirus infection induces vascular injury in the rat. *Atherosclerosis* 1992; **93**(1–2): 41–52.
- 153. Sorlie PD, Adam E, Melnick SL, Folsom A, Skelton T, Chambless LE, et al. Cytomegalovirus/herpesvirus and carotid atherosclerosis: the ARIC Study. J Med Virol 1994; 42(1): 33–7.

- 154. Zhu J, Quyyumi AA, Norman JE, Costello R, Csako G, Epstein SE. The possible role of hepatitis A virus in the pathogenesis of atherosclerosis. *J Infect Dis* 2000; **182**(6): 1583–7.
- 155. Epstein SE, Zhu J, Burnett MS, Zhou YF, Vercellotti G, Hajjar D. Infection and atherosclerosis: potential roles of pathogen burden and molecular mimicry. *Arterioscler Thromb Vasc Biol* 2000; **20**(6): 1417–20.
- 156. Heinemann M, Susa M, Simnacher U, Marre R, Essig A. Growth of Chlamydia pneumoniae induces cytokine production and expression of CD14 in a human monocytic cell line. *Infect Immun* 1996; **64**(11): 4872–5.
- 157. Kaukoranta-Tolvanen SS, Ronni T, Leinonen M, Saikku P, Laitinen K. Expression of adhesion molecules on endothelial cells stimulated by Chlamydia pneumoniae. *Microb Pathog* 1996; **21**(5): 407–11.
- 158. Kol A, Bourcier T, Lichtman AH, Libby P. Chlamydial and human heat shock protein 60s activate human vascular endothelium, smooth muscle cells, and macrophages. *J Clin Invest* 1999; **103**(4): 571–7.
- 159. Kol A, Sukhova GK, Lichtman AH, Libby P. Chlamydial heat shock protein 60 localizes in human atheroma and regulates macrophage tumor necrosis factor-alpha and matrix metalloproteinase expression. *Circulation* 1998; **98**(4): 300–7.
- 160. Kalayoglu MV, Libby P, Byrne GI. Chlamydia pneumoniae as an emerging risk factor in cardiovascular disease. *Jama* 2002; **288**(21): 2724–31.
- 161. Weiss SM, Roblin PM, Gaydos CA, Cummings P, Patton DL, Schulhoff N, et al. Failure to detect Chlamydia

pneumoniae in coronary atheromas of patients undergoing atherectomy. *J Infect Dis* 1996; **173**(4): 957–62.

- 162. Hahn DL, Golubjatnikov R. Smoking is a potential confounder of the Chlamydia pneumoniae-coronary artery disease association. *Arterioscler Thromb* 1992; **12**(8): 945–7.
- 163. Ridker PM, Kundsin RB, Stampfer MJ, Poulin S, Hennekens CH. Prospective study of Chlamydia pneumoniae IgG seropositivity and risks of future myocardial infarction. *Circulation* 1999; **99**(9): 1161–4.
- 164. Stone AFM, Mendall MA, Kaski J-C, Edger TM, Risley P, Poloniecki J, et al. Effect of treatment for Chlamydia pneumoniae and Helicobacter pylori on markers of inflammation and cardiac events in patients with acute coronary syndromes: South Thames Trial of Antibiotics in Myocardial Infarction and Unstable Angina (STAMINA). *Circulation* 2002; **106**(10): 1219–23.
- 165. Epstein SE, Zhu J, Najafi AH, Burnett MS. Insights into the role of infection in atherogenesis and in plaque rupture. *Circulation* 2009; 119(24): 3133–41.
- 166. Eliasziw M, Streifler JY, Fox AJ, Hachinski VC, Ferguson GG, Barnett HJ. Significance of plaque ulceration in symptomatic patients with high-grade carotid stenosis. North American Symptomatic Carotid Endarterectomy Trial. *Stroke* 1994; 25(2): 304–8.
- 167. Rothwell PM, Salinas R, Ferrando LA, Slattery J, Warlow CP. Does the angiographic appearance of a carotid stenosis predict the risk of stroke independently of the degree of stenosis? *Clin Radiol* 1995; **50**(12): 830–3.

- 168. Grønholdt ML. Ultrasound and lipoproteins as predictors of lipid-rich, rupture-prone plaques in the carotid artery. *Arterioscler Thromb Vasc Biol* 1999; **19**(1): 2–13.
- 169. Ehara S, Kobayashi Y, Yoshiyama M, Shimada K, Shimada Y, Fukuda D, et al. Spotty calcification typifies the culprit plaque in patients with acute myocardial infarction: an intravascular ultrasound study. *Circulation* 2004; **110**(22): 3424–9.
- 170. Nair A, Kuban BD, Tuzcu EM, Schoenhagen P, Nissen SE, Vince DG. Coronary plaque classification with intravascular ultrasound radiofrequency data analysis. *Circulation* 2002; **106**(17): 2200–6.
- 171. Hong M-K, Mintz GS, Lee CW, Lee J-W, Park J-H, Park D-W, et al. A three-vessel virtual histology intravascular ultrasound analysis of frequency and distribution of thin-cap fibroatheromas in patients with acute coronary syndrome or stable angina pectoris. *The American journal of cardiology* 2008; **101**(5): 568–72.
- 172. Jang I-K, Tearney GJ, MacNeill B, Takano M, Moselewski F, Iftima N, et al. In vivo characterization of coronary atherosclerotic plaque by use of optical coherence tomography. *Circulation* 2005; **111**(12): 1551–5.
- 173. Tearney GJ, Waxman S, Shishkov M, Vakoc BJ, Suter MJ, Freilich MI, et al. Three-dimensional coronary artery microscopy by intracoronary optical frequency domain imaging. *JACC Cardiovasc Imaging* 2008; 1(6): 752–61.
- 174. Casscells W, Hathorn B, David M, Krabach T, Vaughn WK, McAllister HA, et al. Thermal detection of cellular infiltrates in living atherosclerotic plaques: possible

implications for plaque rupture and thrombosis. *Lancet* 1996; **347**(9013): 1447–51.

- 175. Stefanadis C, Diamantopoulos L, Vlachopoulos C, Tsiamis E, Dernellis J, Toutouzas K, et al. Thermal heterogeneity within human atherosclerotic coronary arteries detected in vivo: A new method of detection by application of a special thermography catheter. *Circulation* 1999; **99**(15): 1965–71.
- 176. Stefanadis C, Toutouzas K, Tsiamis E, Stratos C, Vavuranakis M, Kallikazaros I, et al. Increased local temperature in human coronary atherosclerotic plaques: an independent predictor of clinical outcome in patients undergoing a percutaneous coronary intervention. *Journal of the American College of Cardiology* 2001; **37**(5): 1277–83.
- 177. Botnar RM, Stuber M, Kissinger KV, Kim WY, Spuentrup E, Manning WJ. Noninvasive coronary vessel wall and plaque imaging with magnetic resonance imaging. *Circulation* 2000; 102(21): 2582–7.
- 178. Kim WY, Danias PG, Stuber M, Flamm SD, Plein S, Nagel E, et al. Coronary magnetic resonance angiography for the detection of coronary stenoses. *N Engl J Med* 2001; 345(26): 1863–9.
- 179. Wasserman BA, Smith WI, Trout HH, Cannon RO, Balaban RS, Arai AE. Carotid artery atherosclerosis: in vivo morphologic characterization with gadolinium-enhanced double-oblique MR imaging initial results. *Radiology* 2002; **223**(2): 566–73.
- 180. Aziz K, Berger K, Claycombe K, Huang R, Patel R, Abela GS. Noninvasive detection and localization of vulnerable plaque and

arterial thrombosis with computed tomography angiography/positron emission tomography. *Circulation* 2008; **117**(16): 2061–70.

- 181. Ridker PM, Buring JE, Shih J, Matias M, Hennekens CH. Prospective study of C-reactive protein and the risk of future cardiovascular events among apparently healthy women. *Circulation* 1998; **98**(8): 731–3.
- 182. Ridker PM, Cushman M, Stampfer MJ, Tracy RP, Hennekens CH. Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *N Engl J Med* 1997; **336**(14): 973–9.
- 183. Kai H, Ikeda H, Yasukawa H, Kai M, Seki Y, Kuwahara F, et al. Peripheral blood levels of matrix metalloproteases-2 and -9 are elevated in patients with acute coronary syndromes. *Journal of the American College of Cardiology* 1998; **32**(2): 368–72.
- 184. Loftus IM, Naylor AR, Bell PR, Thompson MM. Plasma MMP-9 – a marker of carotid plaque instability. *Eur J Vasc Endovasc Surg* 2001; 21(1): 17–21.
- 185. Blankenberg S, Rupprecht HJ, Poirier O, Bickel C, Smieja M, Hafner G, et al. Plasma concentrations and genetic variation of matrix metalloproteinase 9 and prognosis of patients with cardiovascular disease. *Circulation* 2003; **107**(12): 1579–85.
- 186. Haim M, Tanne D, Boyko V, Reshef T, Goldbourt U, Leor J, et al. Soluble intercellular adhesion molecule-1 and long-term risk of acute coronary events in patients with chronic coronary heart disease. Data from the Bezafibrate Infarction Prevention (BIP) Study. *Journal of the*

American College of Cardiology 2002; **39**(7): 1133–8.

- 187. Koenig W, Khuseyinova N. Biomarkers of atherosclerotic plaque instability and rupture. *Arterioscler Thromb Vasc Biol* 2007; 27(1): 15–26.
- 188. Sever PS, Dahlöf B, Poulter NR, Wedel H, Beevers G, Caulfield M, et al. Prevention of coronary and stroke events with atorvastatin in hypertensive patients who have average or lower-than-average cholesterol concentrations, in the Anglo-Scandinavian Cardiac Outcomes Trial--Lipid Lowering Arm (ASCOT-LLA): a multicentre randomised controlled trial. *Lancet* 2003; **361**(9364): 1149–58.
- 189. Group HPSC. MRC/BHF Heart Protection Study of cholesterol lowering with simvastatin in 20,536 high-risk individuals: a randomised placebo-controlled trial. *Lancet* 2002; **360**(9326): 7–22.
- 190. Bellosta S, Via D, Canavesi M, Pfister P, Fumagalli R, Paoletti R, et al. HMG-CoA reductase inhibitors reduce MMP-9 secretion by macrophages. *Arterioscler Thromb Vasc Biol* 1998; **18**(11): 1671–8.
- 191. Scalia R, Gooszen ME, Jones SP, Hoffmeyer M, Rimmer DM, Trocha SD, et al. Simvastatin exerts both anti-inflammatory and cardioprotective effects in apolipoprotein E-deficient mice. *Circulation* 2001; **103**(21): 2598–603.
- 192. Endres M, Laufs U, Huang Z, Nakamura T, Huang P, Moskowitz MA, et al. Stroke protection by 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase inhibitors mediated by endothelial nitric oxide synthase. *Proc Natl Acad Sci USA* 1998; **95**(15): 8880–5.

- 193. Shovman O, Levy Y, Gilburd B, Shoenfeld Y. Antiinflammatory and immunomodulatory properties of statins. *Immunol Res* 2002; **25**(3): 271–85.
- 194. Luan Z, Chase AJ, Newby AC. Statins inhibit secretion of metalloproteinases-1, -2, -3, and -9 from vascular smooth muscle cells and macrophages. *Arterioscler Thromb Vasc Biol* 2003; 23(5): 769–75.
- 195. Crisby M, Nordin-Fredriksson G, Shah PK, Yano J, Zhu J, Nilsson J. Pravastatin treatment increases collagen content and decreases lipid content, inflammation, metalloproteinases, and cell death in human carotid plaques: implications for plaque stabilization. *Circulation* 2001; **103**(7): 926–33.
- 196. Ridker PM, Rifai N, Lowenthal SP. Rapid reduction in C-reactive protein with cerivastatin among 785 patients with primary hypercholesterolemia. *Circulation* 2001; **103**(9): 1191–3.
- 197. Aikawa M, Rabkin E, Sugiyama S, Voglic SJ, Fukumoto Y, Furukawa Y, et al. An HMG-CoA reductase inhibitor, cerivastatin, suppresses growth of macrophages expressing matrix metalloproteinases and tissue factor in vivo and in vitro. *Circulation* 2001; **103**(2): 276–83.
- 198. Bourcier T, Libby P. HMG CoA reductase inhibitors reduce plasminogen activator inhibitor-1 expression by human vascular smooth muscle and endothelial cells. *Arterioscler Thromb Vasc Biol* 2000; 20(2): 556–62.
- 199. Libby P, Aikawa M. Effects of statins in reducing thrombotic risk and modulating plaque vulnerability. *Clin Cardiol* 2003; **26**(1 Suppl 1): I11–4.

- 200. Rouis M, Adamy C, Duverger N, Lesnik P, Horellou P, Moreau M, et al. Adenovirus-mediated overexpression of tissue inhibitor of metalloproteinase-1 reduces atherosclerotic lesions in apolipoprotein E-deficient mice. *Circulation* 1999; **100**(5): 533–40.
- 201. Hu Y, Baker AH, Zou Y, Newby AC, Xu Q. Local gene transfer of tissue inhibitor of metalloproteinase-2 influences vein graft remodeling in a mouse model. *Arterioscler Thromb Vasc Biol* 2001; **21**(8): 1275–80.
- 202. Schwartz MA, Venkataraman S, Ghaffari MA, Libby A, Mookhtiar KA, Mallya SK, et al. Inhibition of human collagenases by sulfur-based substrate analogs. *Biochem Biophys Res Commun* 1991; **176**(1): 173–9.
- 203. Watson SA, Morris TM, Parsons SL, Steele RJ, Brown PD. Therapeutic effect of the matrix metalloproteinase inhibitor, batimastat, in a human colorectal cancer ascites model. *Br J Cancer* 1996; 74(9): 1354–8.
- 204. Bigatel DA, Elmore JR, Carey DJ, Cizmeci-Smith G, Franklin DP, Youkey JR. The matrix metalloproteinase inhibitor BB-94 limits expansion of experimental abdominal aortic aneurysms. *J Vasc Surg* 1999; 29(1): 130–8; discussion 38–9.
- 205. Porter KE, Loftus IM, Peterson M, Bell PR, London NJ, Thompson MM. Marimastat inhibits neointimal thickening in a model of human vein graft stenosis. *Br J Surg* 1998; **85**(10): 1373–7.
- 206. Treharne GD, Boyle JR, Goodall S, Loftus IM, Bell PR, Thompson MM. Marimastat inhibits elastin degradation and matrix metalloproteinase 2 activity in a model of aneurysm disease. *Br J Surg* 1999; **86**(8): 1053–8.

- 207. Talbot DC, Brown PD. Experimental and clinical studies on the use of matrix metalloproteinase inhibitors for the treatment of cancer. *Eur J Cancer* 1996; **32A**(14): 2528–33.
- 208. Levitt NC, Eskens FA, O'Byrne KJ, Propper DJ, Denis LJ, Owen SJ, et al. Phase I and pharmacological study of the oral matrix metalloproteinase inhibitor, MMI270 (CGS27023A), in patients with advanced solid cancer. *Clin Cancer Res* 2001; 7(7): 1912–22.
- 209. Johnson JL, Fritsche-Danielson R, Behrendt M, Westin-Eriksson A, Wennbo H, Herslof M, et al. Effect of broad-spectrum matrix metalloproteinase inhibition on atherosclerotic plaque stability. *Cardiovasc Res* 2006; **71**(3): 586–95.
- 210. Greenwald RA, Golub LM, Ramamurthy NS, Chowdhury M, Moak SA, Sorsa T. In vitro sensitivity of the three mammalian collagenases to tetracycline inhibition: relationship to bone and cartilage degradation. *Bone* 1998; **22**(1): 33–8.
- 211. Smith GN, Yu LP, Brandt KD, Capello WN. Oral administration of doxycycline reduces collagenase and gelatinase activities in extracts of human osteoarthritic cartilage. *J Rheumatol* 1998; **25**(3): 532–5.
- 212. Loftus IM, Porter K, Peterson M, Boyle J, London NJ, Bell PR, et al. MMP inhibition reduces intimal hyperplasia in a human vein graft stenosis model. *Ann N Y Acad Sci* 1999; **878**: 547–50.
- 213. Petrinec D, Liao S, Holmes DR, Reilly JM, Parks WC, Thompson RW. Doxycycline inhibition of aneurysmal degeneration in an elastase-induced rat model of abdominal aortic aneurysm: preservation of aortic elastin associated with suppressed production of 92 kD

gelatinase. J Vasc Surg 1996; 23(2): 336–46.

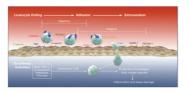
- 214. Thompson RW, Baxter BT. MMP inhibition in abdominal aortic aneurysms. Rationale for a prospective randomized clinical trial. *Ann NY Acad Sci* 1999; **878**: 159–78.
- 215. Axisa B, Loftus IM, Naylor AR, Goodall S, Jones L, Bell PRF, et al. Prospective, randomized, double-blind trial investigating the effect of doxycycline on matrix metalloproteinase expression within atherosclerotic carotid plaques. *Stroke* 2002; **33**(12): 2858–64.
- 216. Baxter BT, Pearce WH, Waltke EA, Littooy FN, Hallett JW, Kent KC, et al. Prolonged administration of doxycycline in patients with small asymptomatic abdominal aortic aneurysms: report of a prospective (Phase II) multicenter study. *J Vasc Surg* 2002; **36**(1): 1–12.
- 217. Miyazaki M, Sakonjo H, Takai S. Anti-atherosclerotic effects of an angiotensin converting enzyme inhibitor and an angiotensin II antagonist in Cynomolgus monkeys fed a high-cholesterol diet. *Br J Pharmacol* 1999; **128**(3): 523–9.
- 218. Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, Dagenais G. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. The Heart Outcomes Prevention Evaluation Study Investigators. N Engl J Med 2000; 342(3): 145–53.
- 219. Tummala PE, Chen XL, Sundell CL, Laursen JB, Hammes CP, Alexander RW, et al. Angiotensin II induces vascular cell adhesion molecule-1 expression in rat vasculature: A potential link between the renin-angiotensin system and

atherosclerosis. *Circulation* 1999; **100**(11): 1223–9.

- 220. Navalkar S, Parthasarathy S, Santanam N, Khan BV. Irbesartan, an angiotensin type 1 receptor inhibitor, regulates markers of inflammation in patients with premature atherosclerosis. *Journal of the American College of Cardiology* 2001; **37**(2): 440–4.
- 221. Hans CP, Zerfaoui M, Naura AS, Catling A, Boulares AH. Differential

effects of PARP inhibition on vascular cell survival and ACAT-1 expression favouring atherosclerotic plaque stability. *Cardiovasc Res* 2008; **78**(3): 429–39.

222. Marczewski MM, Postula M, Kosior D. Novel antiplatelet agents in the prevention of cardiovascular complications – focus on ticagrelor. *Vasc Health Risk Manag* 2010; 6: 419–29.



Cover diagram by David Heinrich of the *Medical Illustration and Media Unit, Flinders Medical Centre*. (See chapter 18)

MECHANISMS OF VASCULAR DISEASE

Edited by Robert Fitridge and Matthew Thompson

Chapter 1: Endothelium Chapter 2: Vascular smooth muscle structure and function Chapter 3: Atherosclerosis Chapter 4: Mechanisms of plaque rupture **Chapter 5**: Current and emerging therapies in atheroprotection **Chapter 6:** Molecular approaches to revascularisation in peripheral vascular disease **Chapter 7:** Biology of restenosis and targets for intervention **Chapter 8:** Vascular arterial haemodynamics **Chapter 9:** Physiological haemostasis **Chapter 10:** Hypercoagulable states **Chapter 11:** Platelets in the pathogenesis of vascular disease and their role as a therapeutic target **Chapter 12**: Pathogenesis of aortic aneurysms Chapter 13: Pharmacological treatment of aneurysms Chapter 14: Aortic dissection and connective tissue disorders Chapter 15: Biomarkers in vascular disease Chapter **16:** Pathophysiology and principles of management of vasculitis and Raynaud's phenomenon Chapter 17: SIRS, sepsis and multiorgan failure **Chapter 18:** Pathophysiology of reperfusion injury **Chapter 19:** Compartment syndrome **Chapter 20:** Pathophysiology of pain Chapter 21: Postamputation pain **Chapter 22:** Treatment of neuropathic pain **Chapter 23:** Principles of wound healing **Chapter 24:** Pathophysiology and principles of varicose veins **Chapter 25:** Chronic venous insufficiency and leg ulceration: Principles and vascular biology Chapter 26: Pathophysiology and principles of management of the diabetic foot **Chapter 27:** Lymphoedema – Principles, genetics and pathophysiology **Chapter 28:** Graft materials past and future Chapter 29: Pathophysiology of vascular graft infections



BARR SMITH PRESS An imprint of The University of Adelaide Press