



# Phenotype characterisation of Phe508del and CFTR knockout cystic fibrosis rats

Alexandra McCarron<sup>1-3</sup>, Patricia Cmielewski<sup>1-3</sup>, Nicole Reyne<sup>1-3</sup>, John Finnie<sup>2,4</sup>, Nathan Rout-Pitt<sup>1-3</sup>, Chantelle McIntyre<sup>2,4</sup>, Fiona Craig<sup>2,4</sup>, David Parsons<sup>1-3</sup>, Martin Donnelley<sup>1-3</sup>

1. Robinson Research Institute, University of Adelaide, South Australia  
2. Adelaide Medical School, University of Adelaide, South Australia  
3. Department of Respiratory & Sleep Medicine, Women's & Children's Hospital, South Australia  
4. SA Pathology, Adelaide, South Australia

## Introduction

- Cystic fibrosis (CF) rats have previously been shown to recapitulate important features of human CF disease (Tuggle et al. 2014, Birket et al. 2018).
- Over the last two years we have developed and bred CF rats with Phe508del and CFTR knockout (KO) genotypes.
- CF rats were primarily generated for the pre-clinical development of airway-directed gene therapies.
- The aim of this study was to characterise the phenotypes of both Phe508del and KO rat models.

## Methods

### CFTR mRNA quantification

- RNA was isolated from lung samples using the Qiagen RNeasy<sup>®</sup> PowerLyzer<sup>®</sup> Tissue & Cells Kit. cDNA was synthesised using the Qiagen QuantiTect reverse transcription kit. CFTR mRNA levels were quantified by qPCR using SYBR<sup>®</sup> green. CFTR expression was normalised to Cyclophilin A mRNA expression.

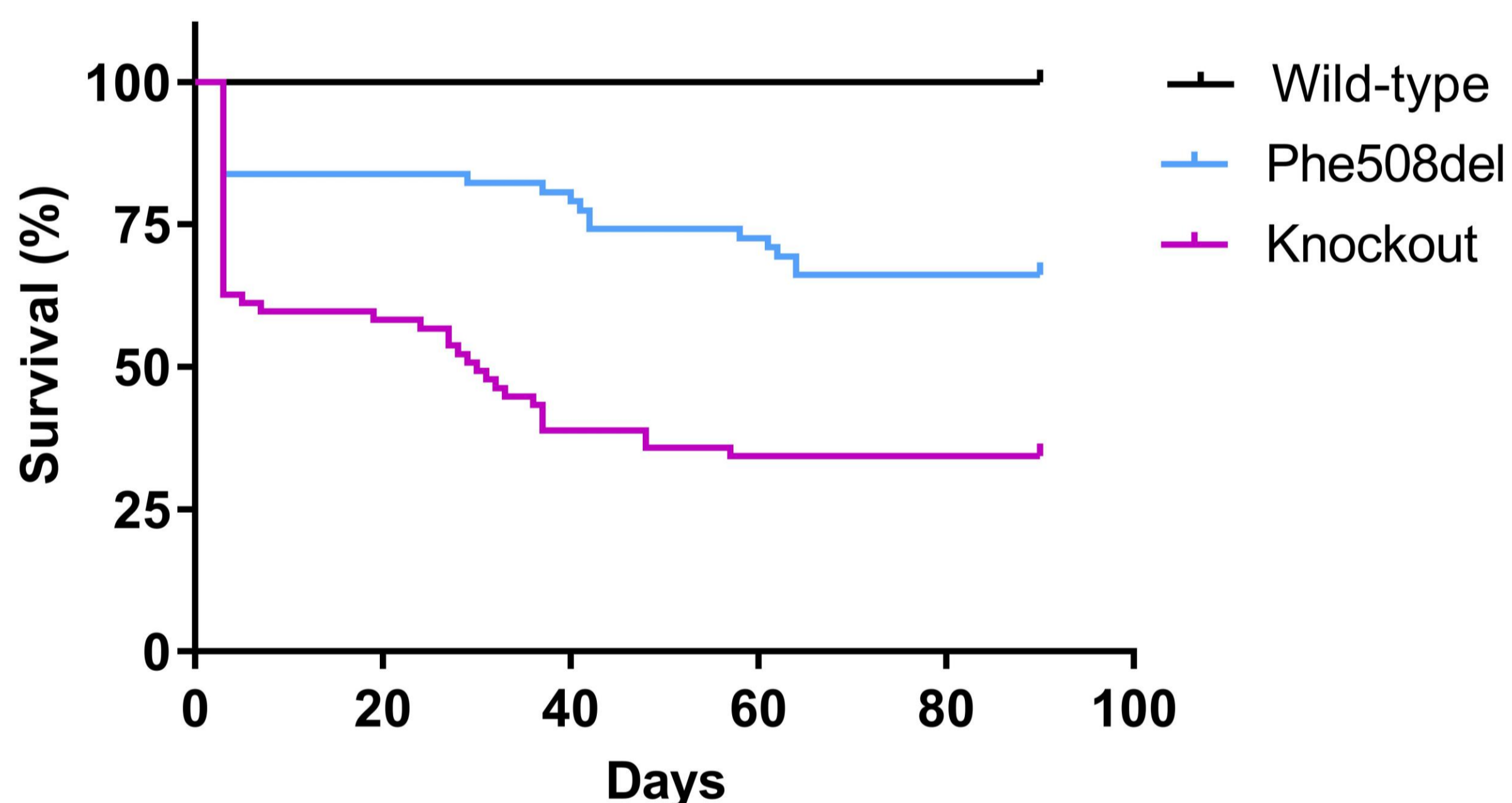
### Nasal potential difference (PD) measurements

- Rats were anaesthetised with domitor:ketamine and non-surgically intubated with a 16 G cannula to permit normal breathing during nasal perfusions of solutions (10-25  $\mu$ l/min). Rats were suspended by their incisors and PD measurements were recorded. Once a plateau of 1-2 minutes was obtained, solutions were then changed. NPD tracings were interpreted by an experienced assessor blinded to the animal genotype.

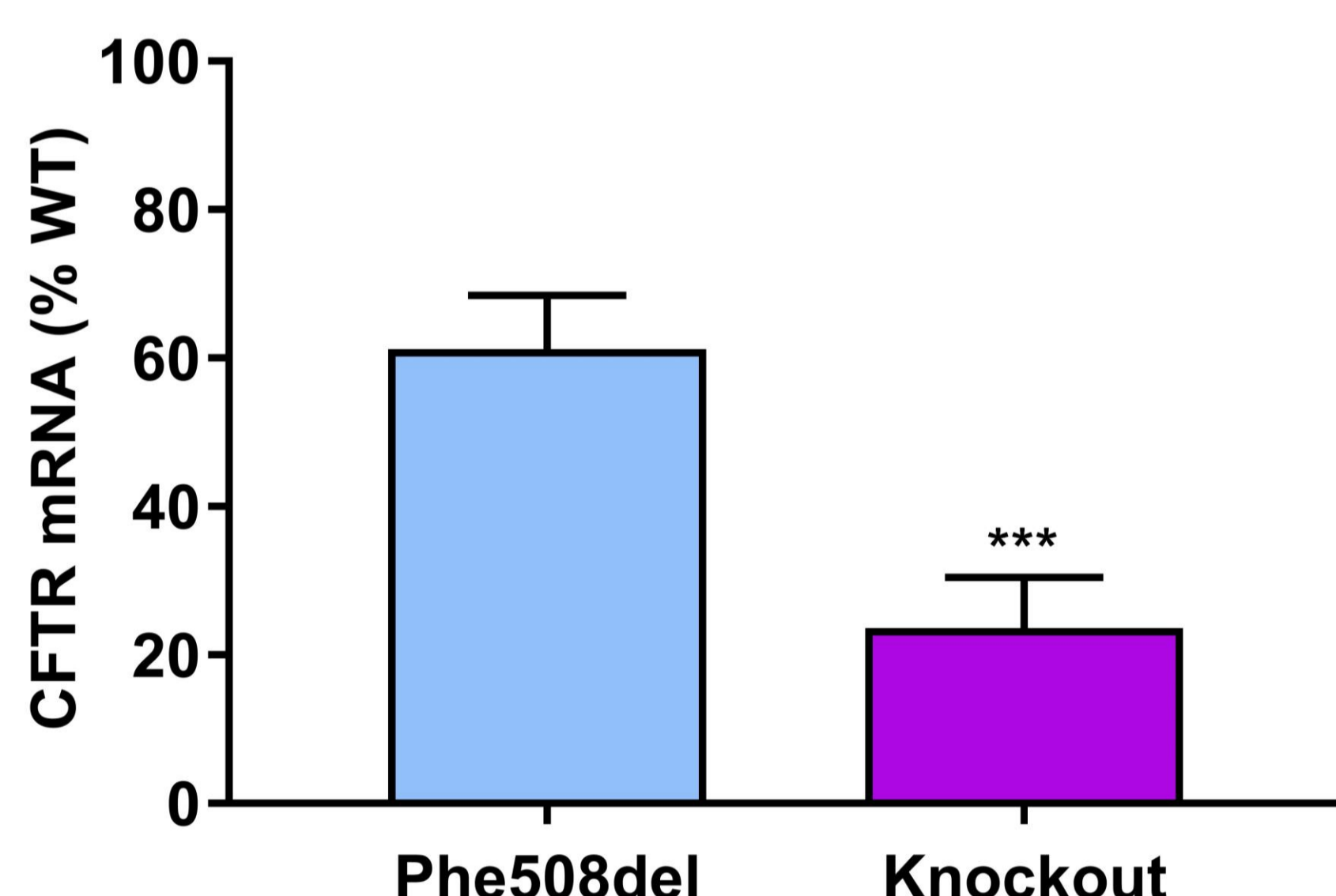
### Histology

- Tissues were fixed in 10% neutral buffered formalin, paraffin embedded, sectioned and stained with hematoxylin and eosin (H&E) or alcian blue periodic acid-Schiff (AB-PAS) for evaluation by a veterinary pathologist.

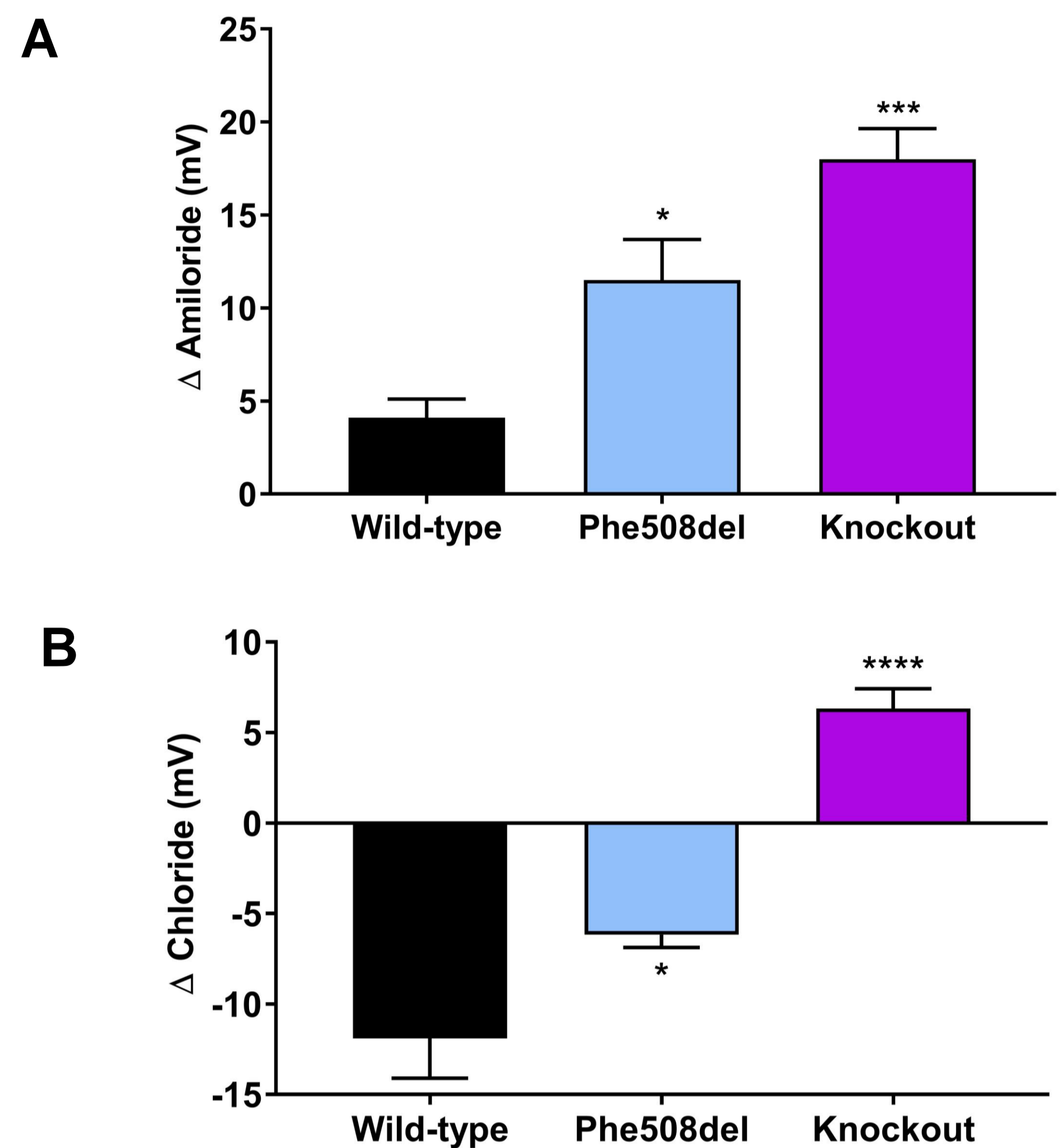
## Results



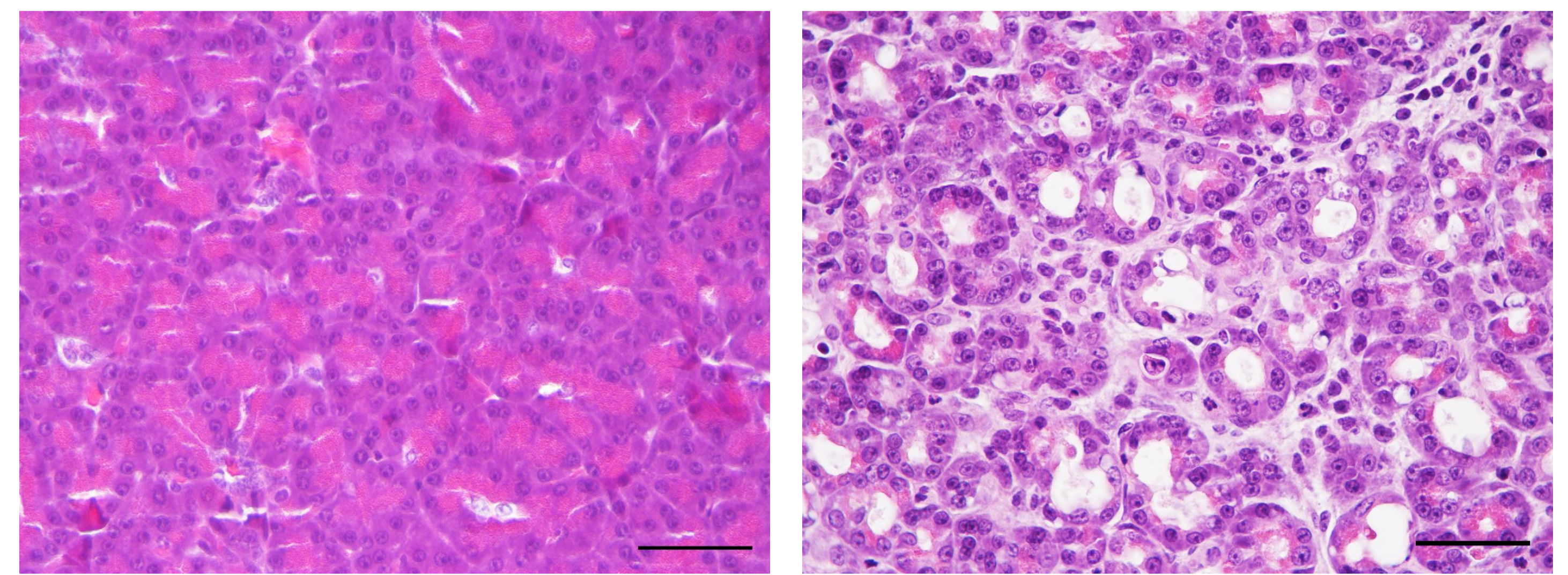
**Figure 1. Survival curve for Phe508del and CFTR KO rats from postnatal day 1 - 90.** Both CF rat models exhibit reduced survival when compared to wild-type rats, with KO rats more severely affected. Early death is attributed to gastrointestinal obstructions and failure to thrive. Statistically significant differences were observed between all groups using Log-Rank test pairwise comparisons (WT vs 508  $p = 7.6 \times 10^{-5}$ , WT vs KO  $p = 3.3 \times 10^{-10}$  and 508 vs KO  $p = 7.6 \times 10^{-5}$ ,  $n = 38-67$  per group).



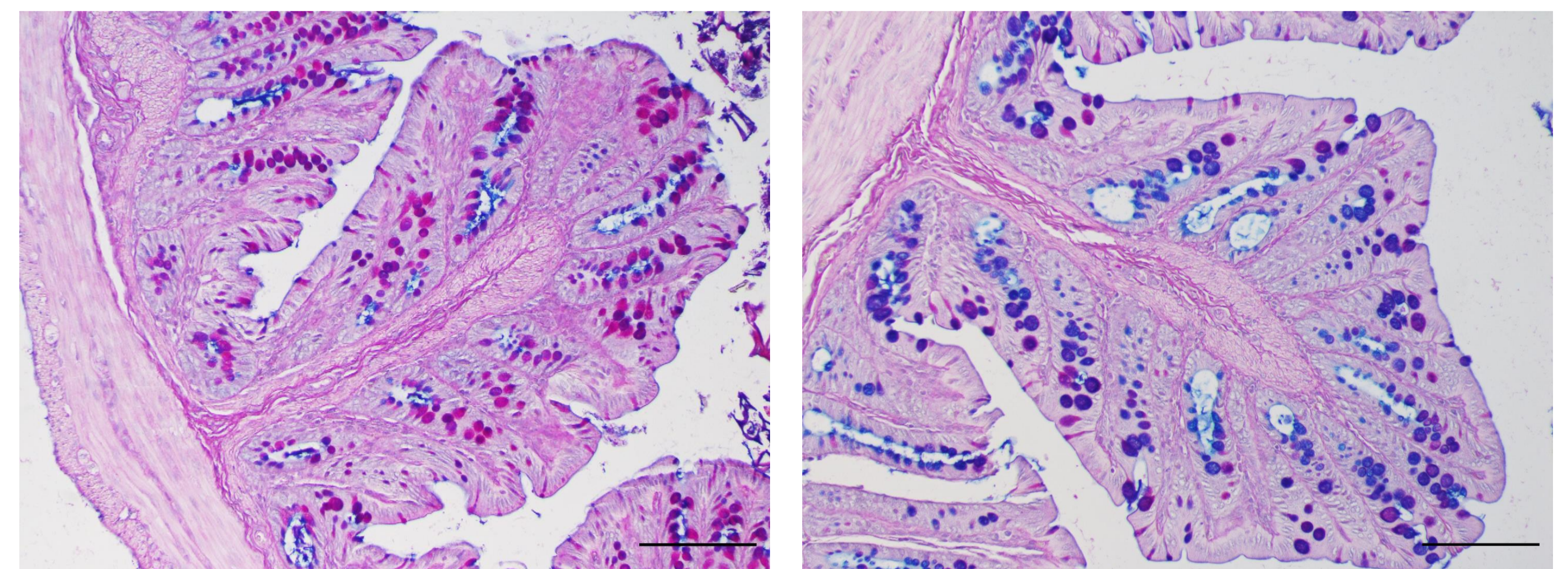
**Figure 2. CFTR mRNA expression levels in Phe508del and CFTR KO rat lungs relative to wild-type.** Phe508del rats had only a small reduction in mRNA levels while KO rats had significantly lower CFTR mRNA expression when compared to wild-type rats (mean with SEM, \*\*\* $p < 0.001$ , one-way ANOVA vs WT,  $n = 7-9$  per group).



**Figure 3. Nasal potential difference measurements in Phe508del and CFTR KO rats.** (A)  $\Delta$ PD response to Krebs-Ringer buffer and amiloride. (B)  $\Delta$ PD response to low chloride Krebs-Ringer buffer and amiloride. When compared to wild-type controls, KO rats demonstrate a classic CF bioelectrical profile while Phe508del rats exhibit a milder phenotype (mean with SEM, \* $p < 0.05$ , \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$ , ANOVA vs WT,  $n = 3-6$  per group).



**Figure 4. H&E stained pancreatic sections from 1 month old Phe508del (left) and CFTR KO (right) rats.** A proportion (~25%) of KO rats demonstrate multi-focal degeneration of the exocrine pancreas, while Phe508del rats exhibit normal pancreas histology. Scale bar = 50  $\mu$ m ( $n = 8$  per group).



**Figure 5. AB-PAS stained large intestine sections from 1 month old Phe508del (left) and CFTR KO (right) rats.** Phe508del and KO rats exhibit distended intestinal crypts that are often dilated with mucins. This intestinal pathology is more pronounced in KO rats. Scale bar = 100  $\mu$ m ( $n = 8$  per group).

## Conclusions

- CF rats demonstrate reduced survival due to gastrointestinal obstructions.
- Lung CFTR mRNA expression is significantly reduced in KO rats when compared to wild-type rats.
- Nasal PD shows a severe CF-like bioelectric phenotype in KO rats and a mild phenotype in Phe508del rats.
- Pancreas and intestine tissues demonstrate abnormal histopathology in both Phe508del and KO rats.