

CYSTIC FIBROSIS MOUSE MODELS DISPLAY AGE DEPENDENT AIRWAY BASAL STEM CELL HYPERPLASIA



Nigel Farrow¹⁻³, Jonathan L McQualter⁴, Martin Donnelley¹⁻³, Patricia Cmielewski¹⁻³, Ivan Bertoncello⁴, and David Parsons¹⁻³

Robinson Research Institute, and 2Center for Stem Cell Research, University of Adelaide, South Australia,

*Department of Respiratory and Sleep Medicine, Women's and Children's Hospital;

The Lung Health Research Centre, Department of Pharmacology, University of Melbourne, Victoria.

Introduction:

We have shown that rare multipotent airway basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay. These basal stem cells in adult mouse airways can be identified and isolated via FACS (EpCAM^{cos}o6-integrin^{cos}CD24^{loos}) and subsequent clonogenic assay.

Methods:

The tracheal airways from 4.5 month and 7 month old normal CF littermates (Het UNC), CFTR knockout (CF UNC), and CFTR gut-corrected (CF FABp) mice were excised and cells disaggregated. FACS-soried cells (CD45^{nog} CD31^{nog} EpCAM^{coo} α6-integrin^{coo} β4-integrin^{coo} CD24^{loo}) from the six groups of mice were then cultured within a matrigel-based dionogenic assay system to quantify the number of basal stem cells present as a percentage of total airway basal cells isolated per trachea.

The lung epithelial colony-forming cell assay:

CD45**©CD31**©EpCAM** CD24** lung calls generate colonies (CFU) comprising cells of both airway and alveolar epithelial lineages when co-cultured in matrigel with Sca-1** EpCAM*** mesenchymal cells and mesenchyme-derived growth factors (Fig 1 & 2).

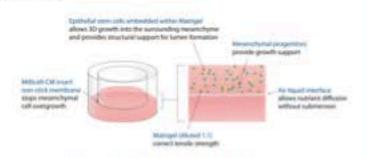


Figure 1: Schematic description of the airway stem cell assay system.

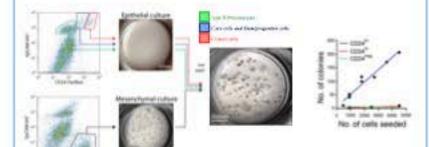


Figure 2: The clonal growth of CD45⁻⁻⁻CD31⁻⁻⁻EpCAM⁻⁻CD24⁻⁻⁻ basal stem cells reveals an obligatory requirement for mesenchymal support. There is a linear relationship between CFU incidence and cells plated. Colony-forming potential is regulated by mesenchyma-derived stimulatory and inhibitory factors.

Upper airway epithelial stem/progenitor cells in mouse models of cystic fibrosis:

The 4.5 month age group displayed a 1.4- fold increase in the number of basal stem cells in the tracheal epithelia of CF UNC mice, compared to normal heterozygous (Het UNC) mice. However, the number of basal stem cells in the CF FABp and Het CF mice were the same at 4.5 months. In contrast, in the 7 month age groups we observed 4.9-fold and 1.9-fold increases in basal stem cells in CF UNC and CF FABp mice respectively, compared to Het CF mice (Fig 3).

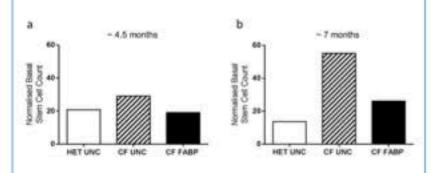


Figure 3: The relative incidence of tracheal airway basel stem cells in Het UNC, CF UNC, and CF EABp mice. A) At 4.5 month age CF UNC mice displayed a 1.4 fold increase in the number of epithelial stem cells compared to the Het UNC mice. B) At 7 month age the CF UNC and CF EABp mice displayed 4.0 and 1.9 fold higher numbers of respiratory epithelial stem cells respectively when compared to the Het UNC controls.

Upper airway epithelial stem/progenitor cell proliferation index:

Sections containing excised tracheae were evaluated for stem/progenitor cell proliferation via immunohistochemistry for a nuclear proliferation antigen, Ki-67. A 2.1 fold increase in proliferating cells was observed in homozygous CF (FABP) mice compared to heterozygous CF (UNC) mice. (Fig 4)

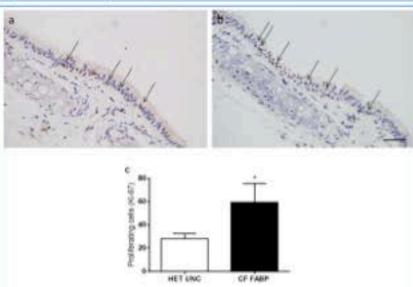


Figure 4: Ki-67 immunohistochemistry of the tracheal airway epithelium from (a) Het UNC and (b) CF FASp mice.

[c] There was a significantly greater number of proliferating calls positive for Ki-67 (black arrows) in the CF FASp mice than the controls (*p < 0.05, 1-test), (scale ber 50 µm)

Summary:

These findings suggest that basal stem cell hyperplasia in the airways of CF mice is not present initially, but may develop as mice age. The increased incidence of basal stem cells in older CF mice suggests there is a progressive increase in the activity of the stem cell compartment, which may contribute to the progressive remodelling of CF airways with age. These findings suggest that early gene therapy correction of CF mouse airways may prevent abnormal hyperplasia of CF airway basal stem cells. If so, hyperplasia of descendant lineages, such as mucin-containing goblet cells, might similarly be beneficially reduced.

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