AIRWAY PRE-TREATMENT ENHANCES MOUSE LUNG LENTIVIRAL REPORTER GENE EXPRESSION

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BACKGROUND:

•Cystic fibrosis (CF) is the most common, fatal genetic disease among the Caucasian population. VSV-G pseudotyped lentiviral (LV) gene vectors have the potential to treat CF long-term, however physiological barriers prevent efficient vector access to ciliated and basal (stem) cells residing on the basement membrane. •We have demonstrated the effectiveness of our lysophosphatidylcholine (LPC) pre-treatment and LV vector dosing technique for producing ciliated and basal cell transduction in mouse nasal airways. •The aim of this experiment was to determine if LPC pre-treatment enhances LV reporter gene transduction in mouse lung conducting airways, the primary target of a CF gene therapy.

RESULTS:

 Dosing was well tolerated, however there was a mild transient respiratory depression at the time of LV delivery.

•There was little to no LacZ transduction in the control group that received the PBS pretreatment, (Fig. 1a) however there was a consistent pattern of strong LacZ transduction of the conducting airways in both LPC treatment groups (Fig. 1b &c).

•Histological staining of lung sections revealed a significant difference in the number of LacZ transduced cells/mm of cartilaginous-associated upper airways from both LPC treatment groups compared to PBS (p<0.01 and p<0.05, ANOVA, 0.1% LPC and 0.3% LPC respectively, Fig. 2).

•The majority of cell transduction occurred in ciliated epithelial cells, with some basal cell transduction also present (Fig. 3).



Figure 1. Example of lung LV-LacZ transduction of conducting airways at 3 months following a) control PBS, b) 0.1% LPC and c) 0.3% LPC. Scale bar 1 mm

CONCLUSION:

LPC pre-treatment prior to LV vector delivery into mouse lungs substantially improves the level of transduction achieved. The LPC pre-treatment groups also showed basal cell transduction that may indicate effective targeting of resident airway stem cells. Doses of LPC lower than the 0.1% level used here should now be trialled to determine the minimally effective conditioning treatment for enhanced lung gene transduction in vivo.







METHODS:

•A 20 µl bolus of a HIV-1 LV vector carrying the LacZ transgene was instilled directly into the trachea of C57BI/6 mice (n=10 per group) via orotracheal intubation, 1 hour after a pre-treatment of either 15µl of PBS (control), 0.1% LPC, or 0.3% LPC.

•LacZ expression was assessed by X-gal staining of the lungs after mice were humanely killed 3 months after dosing.

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